PRELIMINARY STUDIES IN THE GENUS STANHOPEA (ORCHIDACEAE)

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ABSTRACT

This preliminary study is an attempt to bring together the knowledge of Taxonomy, Morphology, Phylogeny and Pollination of the species preparatory to a formal revision of the genus. New information, gathered by population study in the natural habitats, appears to be highly significant to the understanding of this complex group.

The distribution of the species is mapped. The phylogenetic relationships of the genus are considered and it is believed that Stanhopea is derived from plants similar to members of the genus Sievekingia. The development of the advanced and complex species from the simpler species is discussed and the origin of certain unique species such as S. tricornis is considered.

Observations of pollination of S. tricornis by Eulaema meriana and of S. bucephalus by Eulaema bomboides are discussed and pollination relationships are indicated as the major means of genetic

isolation of the two species.

Two highly variable populations were studied in Ecuador. The advanced and variable complexes in Stanhopea are very successful both in area of range and density of population. It is felt that this population structure suppresses speciation as a process, though variability is maintained.

Various nomenclatural problems are mentioned and a key to the species is included. A list of the recognized species is given with their synonyms, and a list is appended of poorly known species which connot as yet he properly placed.

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To say that the genus Stanhopea is poorly understood is an understatement. Rather it is in a state of taxonomic chaos. Before we can say that a genus is understood it is expected that an individual plant, taken at random from a population of similar plants, can be fitted into a species concept. That is to say, it should be possible for a trained botanist to determine whether an individual belongs to a known species or is new and unknown. At the present time the only way to deal with most individual plants of this genus is to name them as new species, thus defeating the whole precept of taxonomy. Throughout its range of distribution Stanhopea is characterized by populations which are extremely variable. The variability within a single population can be so extreme as to appear as a hybrid swarm in which are represented many kinds of Stanhopea hitherto considered as different species.

Because of the plant to plant variability within a single population, a great number of species have been described from single specimens growing in European greenhouses. Often the origin of these plants was not even known. Only the fact that they were different from the other known specimens was taken into consideration and that difference was considered sufficient reason for a new species description. It has been estimated that at least 500 names have been proposed in Stanbopea (Ames and Correll, 1953). This estimate is high, actually only slightly over 100 names have been published. Recent works have reduced that number to between 25 and 50 which are considered as even reasonably valid.

One of the problems involved in the classification of this group is that the flowers are large and fleshy and of intricate structure. They do not preserve well as herbarium specimens and many of the structures of the lip in particular are destroyed or altered in drying. The flowers are not long lasting and seldom persist on the plant for more than three or four days. They are therefore difficult to study

in the field since not many flowers can be seen at any one time within a population. Their period of flowering is often extended over a period of from five to seven months and a plant which is not large will generally flower only once in a given year.

Since they are difficult to study in the herbarium it has been suggested that the only way to straighten out the taxonomic confusion in the genus would be by persistent field work. Our work bears out this suggestion and we do not as yet feel qualified to publish a revision of the genus even though we have studied a considerable number of large populations. We have also studied most of the available herbarium material, but find it hopelessly unrewarding. Such large fleshy flowers as are found in this group do not, as a general rule, make good specimens when crushed and dried. Even after boiling, the critical characters are often lost and determination is difficult. Our experience indicates that dried specimens made from the same plants on different occasions are so altered in drying that they can easily be given separate species designations. This situation is particularly true in certain of the critical species groups which are separated on small differences in structure of the fleshy parts of the lip.

Perhaps all is not as hopeless as it appears on the surface. A number of workers have made limited regional studies for orchid floras. A survey of their work reveals that though they do not all agree, they do not seriously disagree in respect to the majority of the species. In fact, some species appear to be remarkably distinct and do not demonstrate significant variation within their populations. Therefore they are given recognition by each of these workers when they occur in regions treated by them.

When the species in which the problems occur are separated from the ones which are distinct we find that the latter group is not a difficult one. Each species has a distinct distribution and to a certain degree a distinct habitat. Within the problem group the species vary tremendously yet they are in many respects remarkably alike, particularly in morphological characteristics. We feel that this is the key to the problem. They are not only remarkably alike, they are alike and only represent variations of a pattern. If these complexes were treated as representing fewer species each of which may vary considerably within a particular population, then the genus probably could be treated taxonomically and could be understood.

Morphology

Let us consider just what combination of characteristics place a plant in the genus Stanhopea. The vegetative characters are surprisingly uniform, so much so that it is nearly impossible to distinguish species without flowers. All are epiphytic or lithophytic. The pseudobulbs are usually large, ovoid, ribbed and unifoliate. The leaves are broad, elliptic-lanceolate, contracted into a petiole at the base and are strongly veined and plicate. The inflorescence consists of a short pendulous raceme with several broad, papery bracts. The flowers are large, long pedicellate and shielded by a papery bract, this often brightly colored and generally similar in aspect to the sepals and petals. Nearly all species produce a strong fragrance, some being agreeable and others not. The sepals are membranaceous and concave, the dorsal sepal is free and erect and the lateral sepals broad, reflexed and connate at



Fig. 1. Distribution of the genus Stanhopea.

the base. The petals in most species are membranaceous, subequal to the dorsal sepal and recurved. The principal characters which separate the species are to be found in the labellum. In general, the lip consists of three parts: the hypochile or globular, concave part nearest the base of the flowers; the mesochile, which is short and often projected on each side to form horns; and the articulated epichile, which is variously shaped but usually ovate. Certain of the species, such as S. ecornuta Lehm. and S. pulla Rchb. f., simply have a saccate, entire lip which would correspond to the hypocile of the more complex forms. These should perhaps be considered as the more primitive forms and later discussion will attempt to show why. The column is rather similar in most of the species and is elongate and rather slender with or without broad lateral wings. The anther is ventral, incumbent and two-celled. The pollen apparatus consists of a lanceolate viscidium which rests on an elongate spine-like rostellum, a stipe which is long and narrow, and the two pollinia which are elongate, quite thin, and recurved. The stigma forms a pocket under the rostellum.

DISTRIBUTION

The genus is distributed from central Mexico through Central America and most of northern South America into southern Brazil The plants are generally found in localities where at least some shade is afforded and where they will rarely

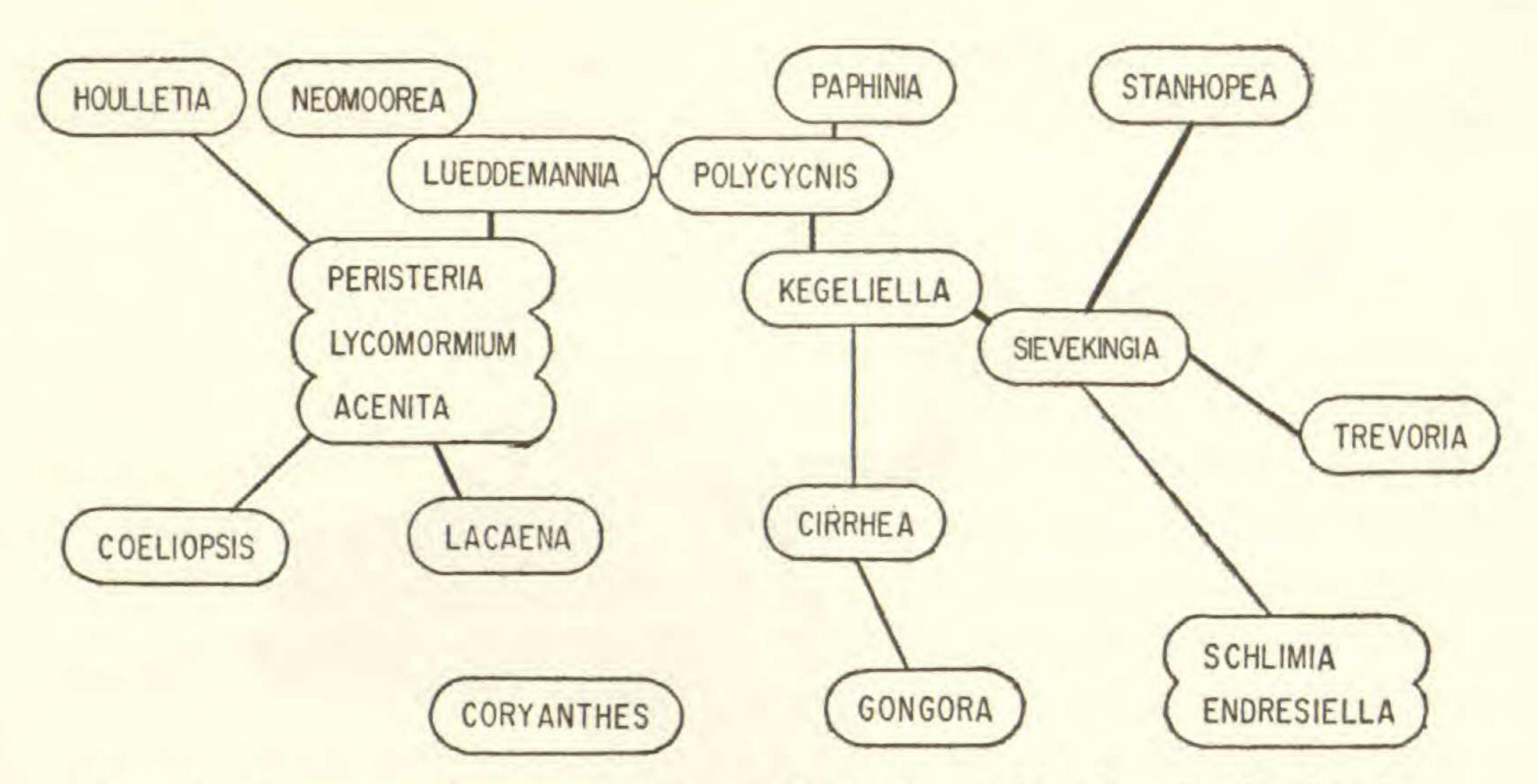


Fig. 2. A diagram of suggested generic relationships within the tribe Stanhopeinae.

become dry. They can withstand prolonged periods of moderate drouth but need considerable water during their growing season.

Figure 1 shows the distribution of the genus. Stanhopea lewisae Ames & Correll occurs in Guatemala and Costa Rica. Stanhopea ecornuta Lindl. occurs in Honduras, Guatemala, Nicaragua and Costa Rica and S. cirrhata Lindl. is found in Nicaragua and Costa Rica. Stanhopea pulla Rchb. f. is found in Costa Rica and Panama. Stanhopea grandiflora Lindl.* extends from Trinidad through Venezuela, northern Brazil and the Amazon drainage region of Colombia, Ecuador and Peru. Stanhopea rodigasiana Hort. and S. reichenbachiana Roezl. ex Rchb. f. are little known species which occur in Colombia. Stanhopea connata Kl. occurs in eastern Ecuador and Peru and S. tricornis Lindl. is restricted to coastal Ecuador and southern Colombia. The remaining species in the genus fall into two major complexes. The first of these groups include S. insignis Frost, S. tigrina Batem., S. martiana Batem. ex Lindl. and S. saccata Lindl. The second group is composed of S. oculata Lindl., S. bucephalus Lindl., S. wardii. Lodd. ex Lindl., and a considerable number of lesser known forms which are quite similar. These two groups occur throughout the range of the genus from Mexico to southern Brazil; however, certain forms are restricted to geographic regions.

PHYLOGENY

The genus Stanhopea was taken by Bentham (1881) as the type of the subtribe and thus the name Stanhopeinae should be used in place of Gongorinae Schltr. (See Dressler and Dodson, 1960). As close allies Stanhopea has the following genera: Sievekingia, Polycycnis, Paphinia, Cirrhea and Gongora and perhaps Peristeria and Houlletia. All of the species within these genera have a labellum which is more

^{*}See the section on Taxonomy for the reason for using this name in place of the commonly used S. eburnea Lindl.

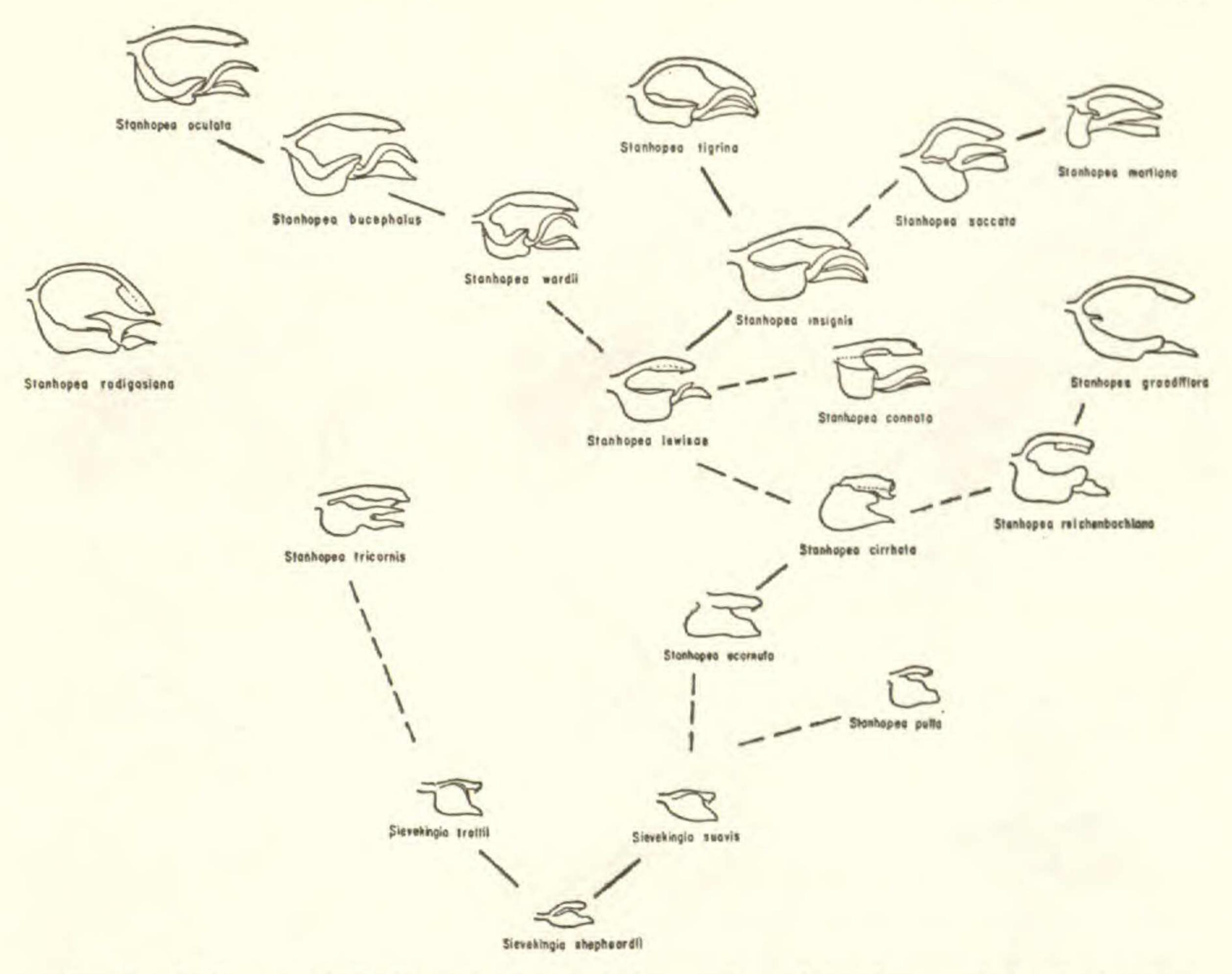


Fig. 3. A diagram of the lip and column of the members of Stanhopea and the closely allied species of Sievekingia suggesting relationships between the species. Solid lines represent close relationships, while dashed lines represent relationships which are more distant or less clear.

or less divided into a hypochile and an epichile. Figure 2 illustrates a possible phylogenetic series demonstrating the relationships of the genera within the Stanhopeinae.

We do not feel that a discussion of the phylogenetic relationships of the other members of the Stanhopeinae would fall within the purpose of this paper and therefore will only discuss those genera which seem to be directly involved with Stanhopea in a stepwise progression from the simple to the complex. The genera placed at the center of the diagram in Figure 2 have a rather simple labellum and as the list radiates outward toward the more advanced genera the lip becomes more complicated.

Figure 3 demonstrates our concepts of the origin and development of the Stanhopeas from the poorly known genus Sievekingia, a genus of about seven species. In general the plants are very similar to small immature Stanhopeas and for this reason are often neglected by collectors. Very few specimens of these plants are in herbaria and in several instances the type is the only specimen in existence. Members of this genus were not discovered until late in the 19th Century and thus did not receive so many generic names as some of the earlier discovered members of the subtribe. Without doubt certain of the species placed in Sievekingia show fewer morphological relationships to each other than do the following genera to each other: Acineta, Lycomormium, Peristeria, Coeliopsis,

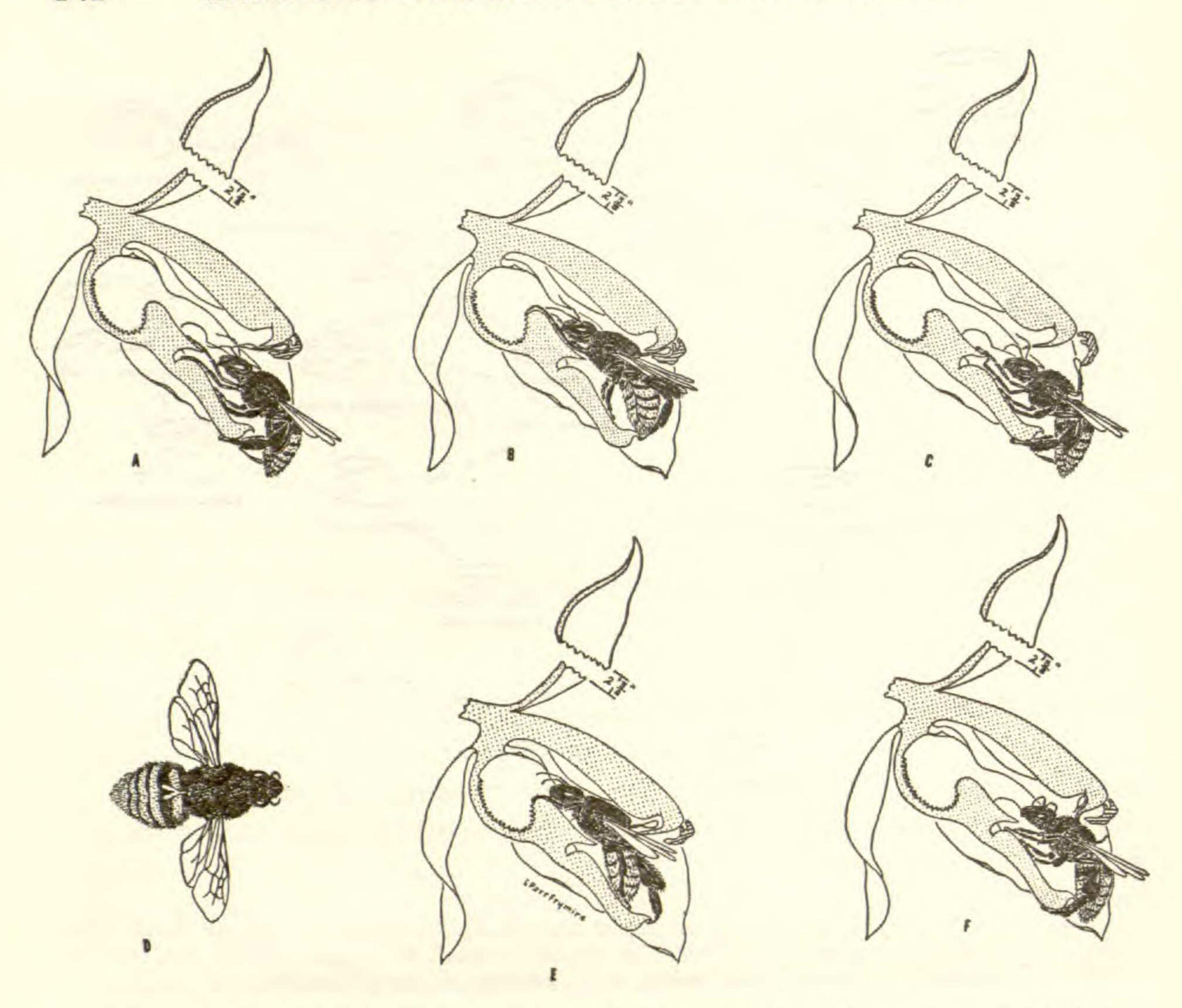


Fig. 4. A diagrammatic representation of the pollination of Stanhopea tricornis Lindl. by Eulaema meriana A. Bee entering the flower. B. Bee backing out of the flower with the spine of the rostellum engaged under the metathorax. C. Bee leaving the flower with the viscidium attached to the metathorax. D. Bee in flight with the pollinarium in place. E. Bee after entering another flower, showing the pollinia inserted in the stigmatic cleft. F. Bee leaving the 2nd flower, showing the pollinia left in the stigmatic cleft.

Neomoorea, Lacaena and Lueddemania. The flowers of the Sievekingias are for the most part less complex than the Stanhopeas. Sievekingia shepheardii Rolfe has a semierect inflorescence with rather simple flowers which have only a slightly lobed, truncate lip with a simple fimbriate callus in the center. This is certainly the most primitive known species at least in respect to the labellum. Sievekingia suavis Rchb. f. and S. peruviana Rolfe have somewhat larger, erect side lobes which form a concave hypochile at the base of the lip. From this point the close relationships of the more primitive Stanhopeas are obvious. It is a simple step from Sievekingia to Stanhopea ecornuta and S. pulla in which the lip is saccate and without the complicated mesochile and epichile of the more advanced Stanhopeas.

Stanhopea cirrhata (Fig. 8) is probably an advancement from a plant similar to S. ecornuta (Fig. 6) or S. pulla (Fig. 7) through the development of lobes along the upper edge of the hypochile. In other respects S. cirrhata is rather similar to the two primitive species except for the narrow, caudate wings formed along each



Fig 5. A diagrammatic representation of the pollination of Stanhopea bucephalus Lindl. by Eulaema bomboides Friese. A. Bee attempting to land on the waxy surface of the hypochile. B. Bee falling down through the flower.

side of the column. This may be the basic form from which the two major branches of Stanhopea have developed. On one side, S. reichenbachiana (Fig. 10) could have been derived from S. cirrhata and could well have been the form from which Stanhopea grandiflora (Fig. 11) developed. Stanhopea grandiflora has a fairly complex lip with short horns produced from the hypochile rather than from the mesochile as in many species.

Stanhopea cirrhata could also be considered as similar to the ancestral form from which S. lewisae (Fig. 13) developed. Stanhopea lewisae has very small, slightly developed horns on the mesochile. Most of the advanced species with well developed horns on the mesochile may well have been derived from S. lewisae or a similar form.

Stanbopea tricornis (Fig. 9) shows little relationship to the other species in the genus. The petals are thick and fleshy and are held parallel to the lip and column. The lip has a hypochile which is only slightly saccate, no true mesochile and a very thick and fleshy epichile. The horns are produced as an extension of the hypochile and a sharp spine-like callus forms a third horn between the two lateral horns in the place of a mesochile. Two possibilities can be postulated for the origin of this unique species. First, it could have developed through evolution from the other known species of primitive Stanbopea as an isolated form. No intermediates are known, however, between this species and the other known Stanhopeas. The second hypothesis may seem a bit far-fetched, but there is some morphological evidence for derivation of S. tricornis from a species of Sievekingia, S. trollii Mansf., recently encountered as sympatric with this Stanbopea. This Sievekingia has petals similar to S. tricornis. The plant is so similar that it doubtless has been passed over by previous collectors (including ourselves) as a young Stanbopea. A colored slide taken of this species, without a scale to indicate size, was at first

determined as a highly floriferous form of S. tricornis. The lip of this Sievekingia is saccate at the base, has two side lobes which flare outward and an epichile very similar to S. tricornis. The callus of the lip is in some respects similar to the center horn of S. tricornis. It seems feasible that S. tricornis may have been derived by way of this species of Sievekingia. Stanhopea tricornis may possibly have been derived as a result of hybridization between Sievekingia trollii or a similar form and some Stanhopea sympatric with it in distribution. Stanhopea bucephalus, S. tricornis and Sievekingia trollii are sympatric along the western slopes of the Andes in Ecuador. If chance pollination of Stanhopea bucephalus by the Sievekingia should occur the resulting intermediate hybrid might well be visited by a pollinator not utilized by either parental species and thereby be stabilized as a species. When an attempt is made to draw an intermediate between the morphological characteristics of the Sievekingia and Stanhopea bucephalus the result is surprisingly similar to the labellum of S. tricornis.

Stanhopea rodigasiana (Fig. 12) is unique in the genus. The strictly oneflowered inflorescence, ovoid, smooth pseudobulbs, bifid rostellum, round viscidium and unique horns on the lip are characters not found in any other Stanbopea. In some respects it suggests a possible relationship to Paphinia, but it is certainly close to Stanhopea. It is surprising to us that this species was never placed in a monotypic genus. Although it is obviously closely related to Stanhopea it is sufficiently different from the other species in the genus to allow any taxonomist with a narrow generic concept to separate it without qualms of conscience. The reason for its not having been so separated is very likely that it was not discovered until after the turn of the century, when more care was being taken in such matters. If it had been discovered 50 years earlier when Peristeria, Acineta, Lycomormium, and other closely allied genera, were being separated, it would surely have been considered as generically distinct. We do not believe that it warrants special distinction nor do we believe that so many genera should be recognized in the Peristeria complex. We hope that through more careful study of the entire subtribe Stanhopeinae we will be able to develop a useful revision of its members.

The above mentioned species of Stanhopea are for the most part easily recognized, have distinct morphological characters and distributional patterns. They present no serious problems to the taxonomist. The remaining species within the genus form the difficult portion. All of them have a more complex labellum than the species previously discussed. The hypochile is saccate and of various sizes but it follows a similar pattern in all of the species. The mesochile is short and inserted on the apex of the hypochile with two lateral, elongate horns of various sizes. The epichile is articulated to the apex of the mesochile and while it may vary in size and proportions, they are all similar. Two major groups can be separated and while we are not yet prepared to present a formal revision of the genus we feel that some designation is necessary in order to be able to group the excessively large number of species names in such a way that they can be discussed with relative ease. We will use the terms "oculata complex" for those plants similar to S. oculata, and "insignis complex" for the plants allied to S. insignis. The "oculata complex" will therefore comprise those concepts which have a rather long narrow or rectangular

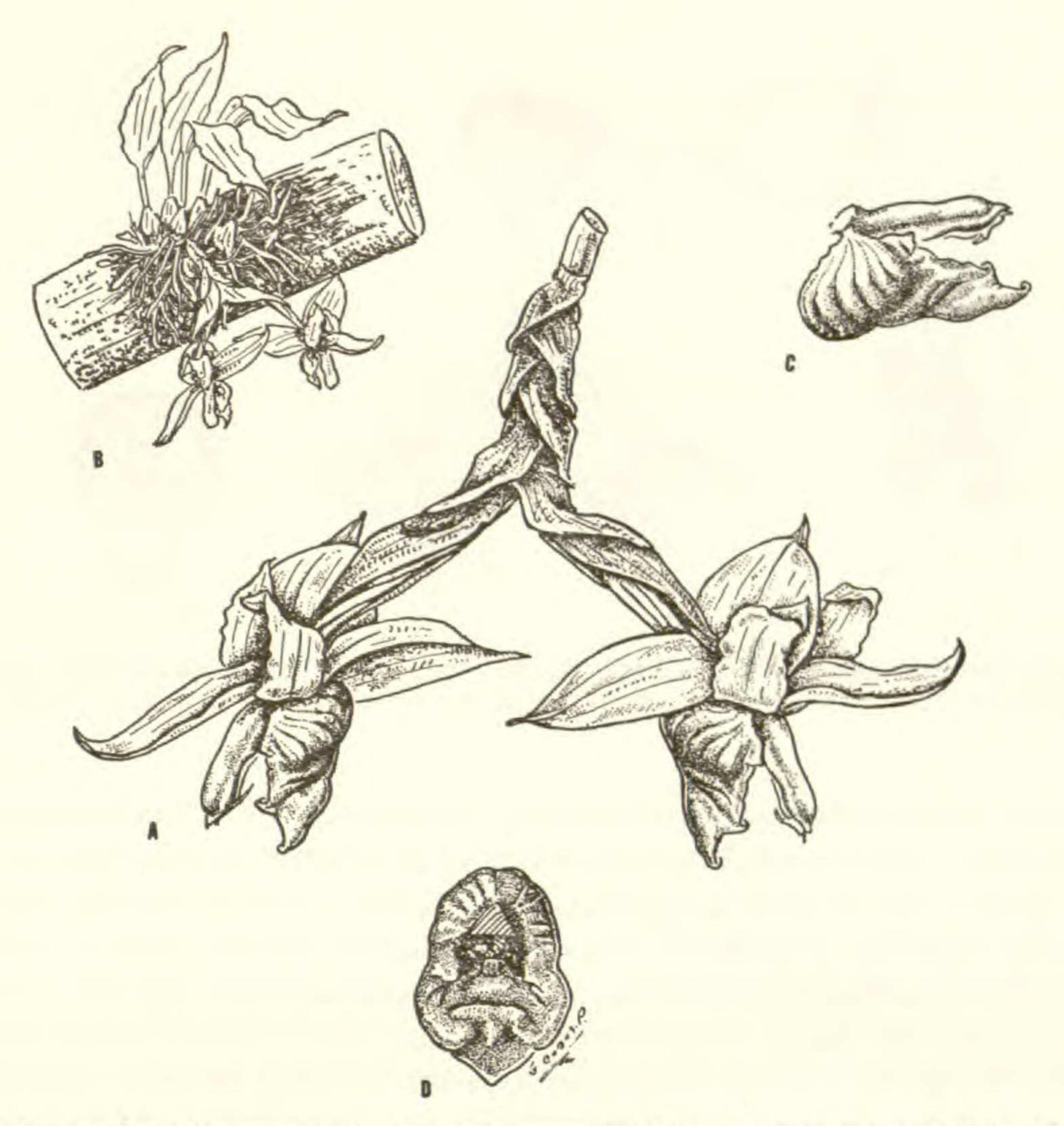


Fig. 6. Stanhopea ecornuta Lem. A. Inflorescence $(\times \frac{1}{3})$. B. Plant habit $(\times \frac{1}{9})$. C. Side view of the lip and column $(\times \frac{1}{2})$. D. Dorsal view of the lip $(\times 1)$. E. Pollinarium $(\times 3)$.

hypochile, large mesochile horns and always have an entire, acute or apiculate epichile. The "oculata complex" comprises such taxa as: S. wardii Lodd. ex Lindl., (Fig. 21), S. oculata (Lodd.) Lindl. (Fig. 19), S. bucephalus Lindl, (Fig. 20), S. haseloviana Rchb. f., S. platyceras Rchb. f., etc. The "insignis complex" will consist of those forms which have a saccate or globular hypochile, large mesochile horns and either entire, acute or three-lobed epichiles. The "insignis complex" comprises such species as S. tigrina Batem. ex Lindl. (Fig. 16), S. martiana Batem. ex Lindl. (Fig. 17), S. insignis Frost (Fig. 15), S. saccata Batem. (Fig. 18) and S. devoniensis Lindl.

Stanhopea quadricornis Lindl., if such a species exists, has a lip which is within the variation pattern of S. oculata with the exception that the base of the thickenings of the margin of the hypochile are projected into short horns. Apparently only one specimen was collected and was named by Lindley. It is quite possible that it was only an aberrant specimen or a hybrid and was not representative of any population.

It seems apparent that Stanhopea is a rather terminal group in evolution, with Coryanthes, a genus often said to be closely allied to Stanhopea, having a derivation

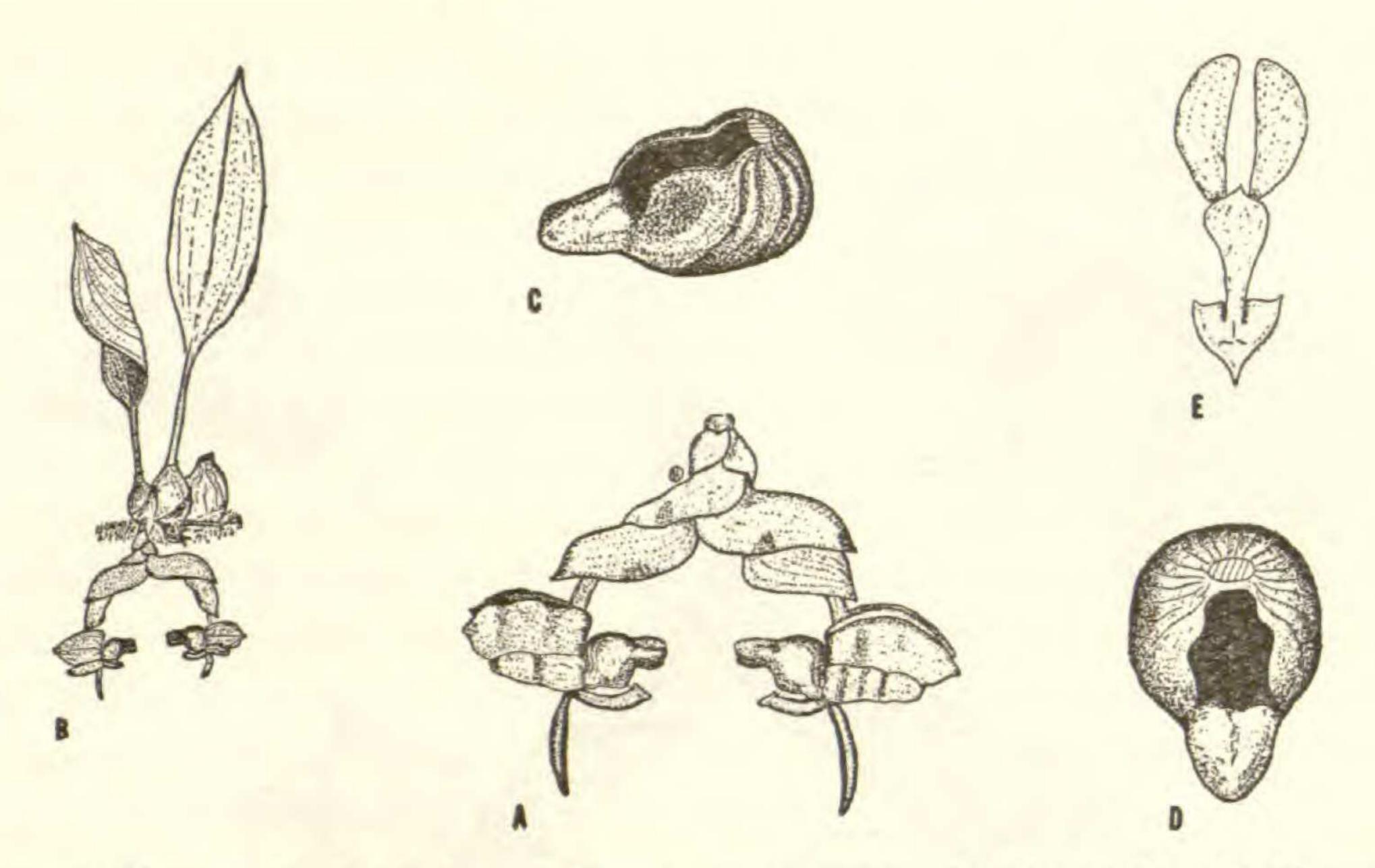


Fig. 7. Stanhopea pulla Rchb. f. A. Inflorescence $(\times \frac{1}{3})$. B. Plant habit $(\times \frac{1}{12})$. C. Side view of the lip $(\times 1)$. D. Dorsal view of the lip $(\times 1)$. E. Pollinarium $(\times 3)$.

from some similar ancestor, but one probably more closely allied to Peristeria than to Stanhopea. Cirrhea and Gongora appear to be the termination of a separate line.

In general, the Orchidaceae appears to be a rather recent but rapidly evolving family in which less extinction is evident than in older families. Such a situation often makes classification difficult due to the intergrading forms, but on the other hand, it makes the tracing of phylogeny easier since closely related groups are still present. We do feel that the Stanhopeas were derived from ancestors very much like some of the species of Sievekingia which are known today. The species of Sievekingia are not particularly successful, as is evident from their extreme rarity, but may be considered as relict forms locally preserved in relatively hospitable habitats. Such forms would very likely become extinct and be removed from the record if they were members of plant families subjected to more stringent environmental conditions.

POLLINATION

This paper is perhaps somewhat premature in one respect, that is, the pollinators and pollination mechanisms are not as yet known for all the species. However it may be many years before all the information can be gathered. We feel that the major worth of the paper lies in its discussion of the problems within the genus.

The pollination mechanisms and pollinators of two species have been well worked out and photographed. Pollination studies of the other species will be described when they are known. We have witnessed the pollination of both Stanhopea tricornis and S. bucephalus in Ecuador.

A future revision of this genus may well be based in large part upon the ideas gained from observations of pollination. Polymorphic genera are often difficult to understand and when confusion is compounded by having highly variable populations, the naming of which has been multiplied on an unrealistic basis, such a genus

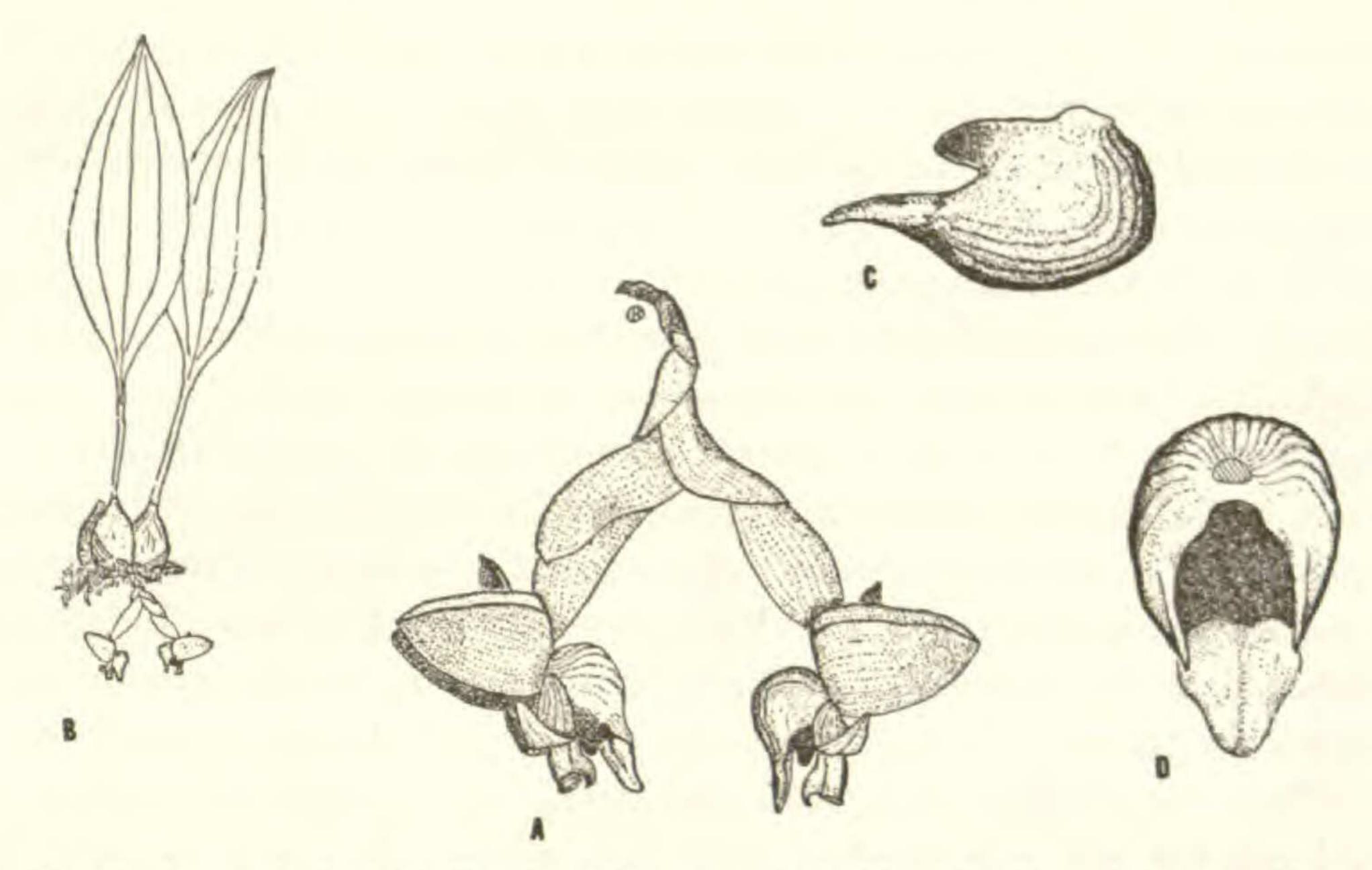


Fig. 8. Stanhopea cirrhata Lindl. A. Inflorescence (X 1/3). B. Plant habit (X 1/17). C. Side view of the lip (X 2/3). D. Dorsal view of the lip (X 2/3).

becomes chaotic. Occasionally new techniques or observations from a different standpoint, can provide flashes of insight for the investigator. The totally different pollination mechanisms of these kinds of Stanhopeas, when observed, immediately and clearly made obvious certain of the reasons for the polymorphism within the genus. The first system is typical of most orchids in which the pollinator simply enters the flower after landing on the expanded labellum. The transfer of pollinia occurs as a result of proper positioning of the column and its sexual apparatus in relation to the size and conformity of the particular pollinator. The majority of the species of *Stanhopea* are of this type. These species are not extremely variable and are easily recognized.

The second is a highly complex system in which the flight characteristics of the bee are used in the pollination of the flower. After landing, the bee is clumsy and has difficulty in regaining balance in flight. The bee, unable to grasp the slick surfaces of the pendant labellum, falls through the flower. This characteristic flight habit of the bee probably has contributed to the evolution of an entirely different system of pollination. Though this kind of flower is distinct, it apparently has been unable to preclude occasional hybrids within the related forms but has simply led to more variation. Color and morphology of the unessential flower parts appear to have been subject to little selection pressure for uniformity, and may vary considerably from plant to plant within a given population. The resulting confusion, caused by attempts to name every variant in cultivation, has not only increased out of all proportion but has tended to make the genus nearly impossible taxonomically.

Stanbopea tricornis occurs frequently in old epiphyte covered cacao trees in the region of Quevedo in central, coastal Ecuador. The species generally succeeds best in fairly dense shade and on a large plant the pendant two-flowered racemes are produced throughout the year. Several racemes may be produced by one pseudobulb.

In late January of 1960 we succeeded in finding a plant which had two open flowers (both had been pollinated) and two mature buds which would open the following day. We removed the plant and carefully carried it to our hotel room. Before dawn of the next morning we found the flowers to be open and emitting a heavy perfume very similar to "Diorissimo" manufactured by the Christian Dior Company of Paris, France. We returned to the cacao plantation at dawn and set the plant in a position where it could be easily photographed. At about 9 a.m. a large male bee, Eulaema meriana (Oliv.) (tribe Euglossini, family Apidae, E. dimidiata is a synonym) came to the flowers, which he proceeded to investigate, apparently searching for the source of the strong fragrance. Meanwhile a second bee of the same species arrived; neither bee seemed familiar with the flower and each searched for some time before discovering the tunnel-like opening, formed by the petals, column and lip, and entering the flower. It might be noted that this is the only species of Stanhopea in which the petals are fleshy and held closely appressed to the column. The bee forced upward past the rostellar spine and entered the inner portion of the flower to the hypochile (see Fig. 4). The bee did not attempt to extract food. In backing out the bee encountered the rostellar spine once again. This time the rostellar spine slid under the posterior projection of the metathorax of the bee, and his struggles dislodged the lance-shaped viscidium of the pollinia, which immediately became cemented to the underside of the metathorax. In further backing out he pulled the pollinia from the anther cap. The bee then flew away. Even if the bee had entered the other flower, pollination could not have been accomplished immediately for the stigmatic pocket of Stanhopea does not open sufficiently for insertion of pollinia until several hours after the pollinia have been removed. The following morning what appeared to be a different bee but one with pollinia attached to his metathorax visited the flowers. The pollinia became lodged in the stigma and were removed from the back of the bee, thus effecting pollination. In total, five instances of pollination of Stanhopea tricornis were observed with six bees, all E. meriana, involved.

Other species of Eulaema were noted in the immediate vicinity and were quite curious about the camera, approaching to examine it, but showed no interest in the Stanhopea tricornis flowers only a foot or so away. These species were E. tropica (Linné) and E. cingulata (Fab.)* both of which are responsible for pollinating numerous other orchids including several species of Catasetum, Cycnoches, Sobralia violacea Lindl., Maxillaria grandiflora Lindl., Aspasia epidendroides Lindl. and Pescatoria wallisii Linden & Rchb. f.

From these observations we can glean the following important points:

- 1. Eulaema meriana appears to be the specific pollinator of S. tricornis.
- 2. Eulaema tropica and E. cingulata are sympatric but showed no interest in S. tricornis.
- 3. Eulaema meriana is attracted to the fragrance emitted by S. tricornis and without prior experience with the flower must search to find the entrance.

^{*} These identifications have been recently supplied by Dr. Krombein of the Smithsonian Institution. These two species were previously known as E. musitans and E. fasciata respectively.

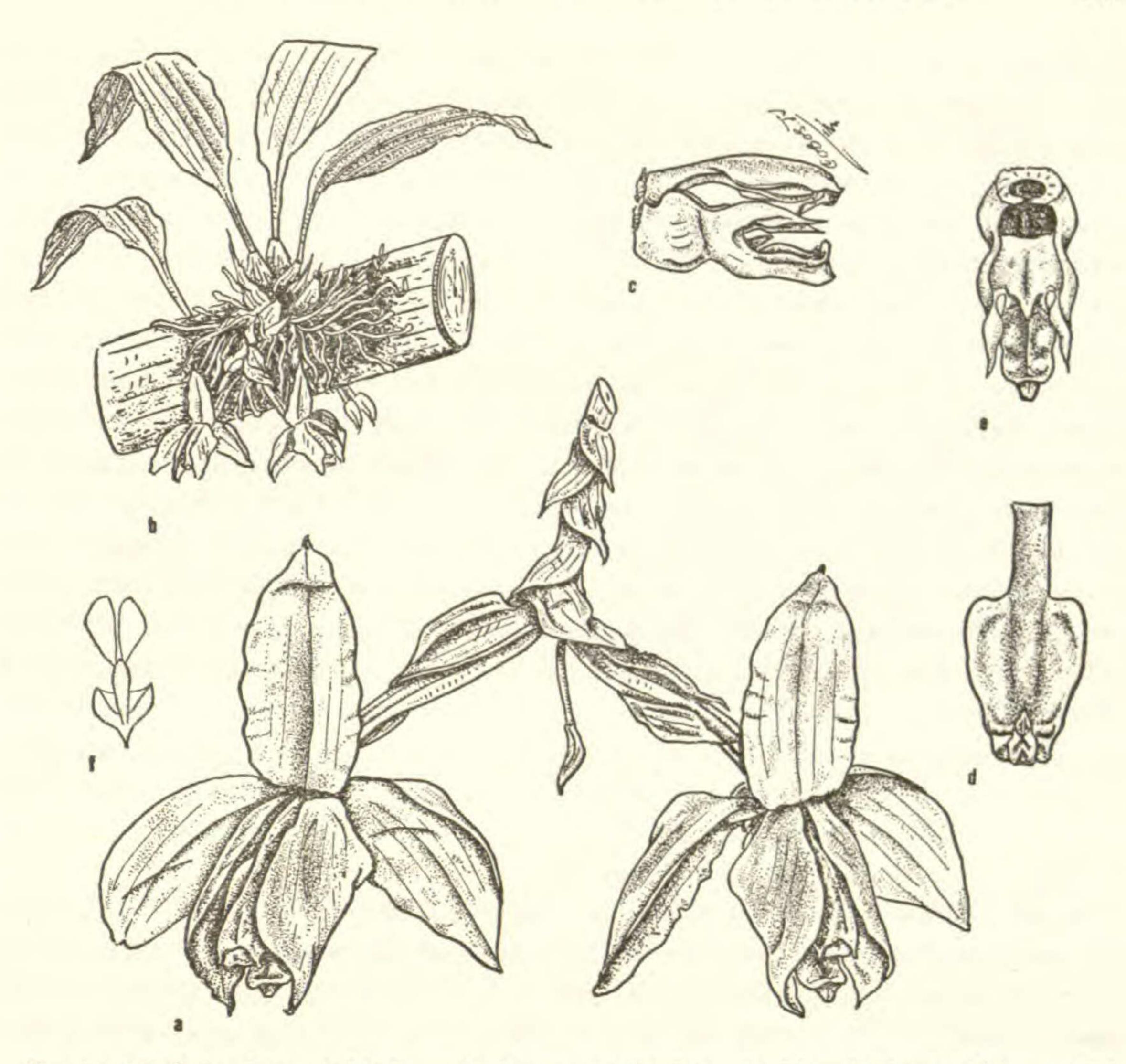


Fig. 9. Stanhopea tricornis Lindl. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{12})$. c. Side view of the lip $(\times \frac{1}{3})$. d. Ventral view of the column $(\times \frac{1}{2})$. e. Dorsal view of the lip $(\times \frac{1}{3})$. F. Pollinarium $(\times 2)$.

Flower color and structure are certainly secondary if even of importance in the actual attraction of the bee.

Flowers of S. tricornis were carefully examined for nectaries or other anatomical structures which would offer food for the bee. None was found. The fragrance is produced in the hypochile; the other parts, when removed from the flower, have little or no fragrance. The inside of the hypochile is covered with long fleshy hairs which the bee might eat but in the flowers entered by E. meriana these hairs did not appear to have been disturbed. It appears then, that the bee is attracted strictly on the basis of fragrance and does not receive food. He apparently does not expect to receive food for he does not lower his tongue, which is so long that it must be lowered and swung forward before a narrow flower is entered. This pattern has been observed in other members of the Catasetinae and Stanhopeinae in which male Eulaemas are attracted by the fragrance and the bees do not appear to receive food of any kind.

In all of these orchids the flowers are generally not long lasting but produce their strong, heady, spicy fragrance in such quantity that they can be detected by the human nose, a notably weak olfactory organ, at distances of more than thirty feet. The bees are usually quite wary when they first come to the flowers but after being around them for some time seem to become drugged for they fly erratically and can be approached easily. Only the male bees of Eulaema appear to be attracted to this type of flower; the females, which are busy with nest building and food gathering to provision the nests, do not approach these flowers. The male bees have never been observed taking part in nest building or care of the nest and seem to leave the nest as soon as possible and live a vagabond life. They secure food from many of the same flowers as the female bees but also visit Stanhopeas, Catasetums, Cycnoches and other flowers which offer only the heady and seemingly intoxicating fragrances. It is possible that the female bees are not attracted to these flowers because they cannot sense them. It is known that female bees of Apis mellifera, the domestic bee, have fewer olfactory pits in their antennae than do the males. The only orchids in which both male and female bees were noted were Sobralia violacea and Maxillaria grandiflora. Sobralia is a rather primitive orchid which does provide nectar for the bee and which is visited by several bees of different families. Maxillaria grandiflora provides starchy hairs at the base of the lip. One morphological feature found in the male bees of the Euglossini which is not present in the females is the presence of chemoreceptive hairs on the tarsi of the front legs. The male bees brush these pads through the scratched surface of the labellum of the orchids which they visit.

Stanhopea bucephalus is pollinated in a somewhat different manner. This variable species is distributed along the western slopes of the Andes from elevations of about 400 meters up to 1200 meters and is to be found on the summits of the coastal mountains where they rise to these elevations. It is sympatric with S. tricornis in the region of Quevedo and the two species appear to be sympatric for at least 60 kilometers to the south.

Pollination of this species was observed during the month of March at Olimpo on the railroad from Guayaquil to Quito. Unlike S. tricornis this species has thin, narrow petals which became retracted and curl up around the dorsal sepal shortly after the flower opens. The horns are not produced as an extension of the hypochile as in S. tricornis but are larger and developed from a definite mesochile. Thus there is left a large open space between the lip and the column at the level of the hypochile. The lower part of the column is effectively surrounded, to form a tunnel, by the horns of the mesochile, the epichile and the wings of the column. The pollinator of this species is also a male Eulaema, E. bomboides Friese, but this one is only about two thirds as large as E. meriana. Eulaema bomboides is a little larger than either E. cingulata or E. tropica.

Eulaema bomboides is attracted by the fragrance of this Stanhopea which is honey-sweet, quite unlike S. tricornis. The bees come to the flower of S. bucephalus but immediately fly into the space between the column and the hypochile and attempt to land on the hypochile, or they land on the side of the hypochile and attempt to crawl around and enter the hypochile to reach the source of the fragrance. The surfaces of all parts of the lip are waxy and extremely slick, so that the bee can not successfully land and hold fast. He falls down and is guided

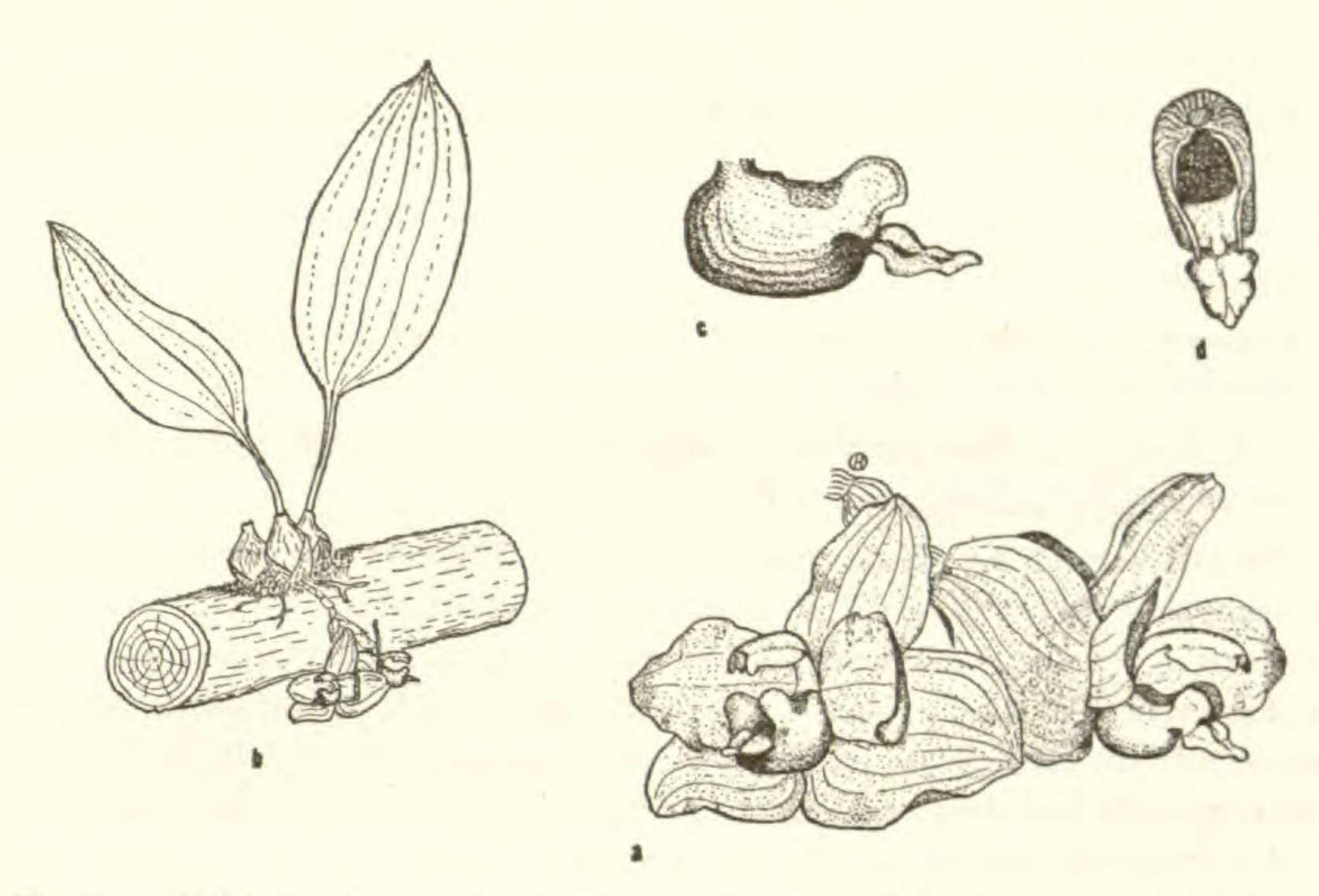


Fig. 10. Stanhopea reichenbachiana Roezl. ex Rchb. f. a. Inflorescence (X 1/3). b. Plant habit (X 1/12). c. Side view of lip (X 1/3). d. Dorsal view of the lip (X 1/3).

by the column and its wings at his back, the horns of the mesochile at his sides and the slick epichile beneath him so that he falls directly through the tunnel. At the lower end of the tunnel, almost at its exit, the rostellar spine with the viscidum of the pollinia on its outer surface projects into the path of the falling bee and is caught under the posterior projection of his thorax as in S. tricornis. The bee hits the rostellar spine with such force that it gives way and allows the bee to fall through. In doing so the viscidium of the pollinia is cemented firmly into place under the thorax (see Fig. 6). The bee is apparently disturbed and tends to fly away but he repeats the process when he encounters another flower. If the flower is older with an open stigma the pollinia are this time placed in the stigmatic pocket during his fall and pollination is effected.

The pollination of S. bucephalus is in many respects quite similar to the pollination of Gongora maculata Lindl. as reported by Allen (1956), with the exception that the Gongora is pollinated by species of Euglossa. Euglossa is a genus of bees which is very closely related to Eulaema and is also responsible for the pollination of Coryanthes (Allen 1950). In Gongora the bee, hanging upside down loses its hold on the slick surfaces of the lip, falls and is guided down the pendant column during his fall with the result that he also makes contact with the viscidium of the pollinia much in the same manner as Eulaema in S. bucephalus.

Several plants of this Stanhopea were brought back to the garden of the Instituto Botánico where quantities of Eulaema tropica and E. cingulata are common and at no time did any of these bees show interest in the flowers. Even when flowering plants of Catasetum were placed in the immediate proximity the Eulaemas ignored the Stanhopea flowers completely.

From these observations we can add the following facts:

- 1. Eulaema bomboides appears to be the specific pollinator of S. bucephalus in the region studied.
- 2. Stanhopea bucephalus is pollinated on an entirely different principle from S. tricornis.
- 3. Eulaema bomboides is attracted by the fragrance of S. bucephalus which is different from the fragrance of S. tricornis.
- 4. If E. bomboides does not visit S. tricornis then S. tricornis and S. bucephalus are effectively isolated genetically.

The last point is by far the most important. Eulaema meriana is too large to fall through S. bucephalus. If E. bomboides is not attracted by the fragrance of S. tricornis then the possibilities of the two species of Stanhopea forming hybrids are effectively nil. This appeared to be the case. Stanhopea tricornis shows very little variability throughout its populations. The sympatric species S. bucephalus is quite variable but shows no characters indicating introgression from S. tricornis.

If S. tricornis evolved in this region with E. meriana as its pollinator and S. bucephalus was dependent upon a different insect pollinator and a different pollination procedure the two would not hybridize. If by accident a bee of E. bomboides did enter S. tricornis and effect a hybridization between the two species of Stanbopea, and it is feasible that this might occur, then the resulting hybrids would probably find no effective pollinator. If such a pollinator did exist then the chances might be better for development of a completely different, stable species with its own characteristics and its own structurally isolated population.

Interestingly enough, perhaps not all Stanhopeas are pollinated by members of the genus Eulaema. The type locality of Euglossa viridissima Friese is given as "at Stanhopea tigrina at Cordoba and Orizaba in Mexico". If this report is correct, and there is no reason to doubt its validity, it indicates that perhaps some of the species are even more widely separated ecologically than are S. tricornis and S. bucephalus. Our observations indicate, at least in Ecuador, that Euglossas and Eulaemas are not mutually attracted to the same orchid flowers. Where they are attracted to orchid species of the same genus (i.e., Eulaema cingulata is attracted to Cycnoches lehmanii and C. ventricosum while Euglossa viridissima is attracted to C. egertonianum) the flowers are extremely distinct morphologically. The principal morphological difference between S. tigrina and S. bucephalus which would be of importance in pollination procedure lies in the extreme closeness of the column and terminal portions of the lip in the former species whereas in the latter these structures are rather open. A large bee like a Eulaema would not be able to fall through S. tigrina due to this constriction of parts. On the other hand a Euglossa would fall through S. bucephalus without touching any of the parts and would be ineffectual in pollination. We feel that the significant factor in this situation does not lie in the morphological form being a deterrent to effective pollination, but rather that the fragrance of the flower does not attract a pollinator which would be ineffective. More likely the morphological structures of the flower have been modified to accommodate the type of pollinator which was attracted to the flower.

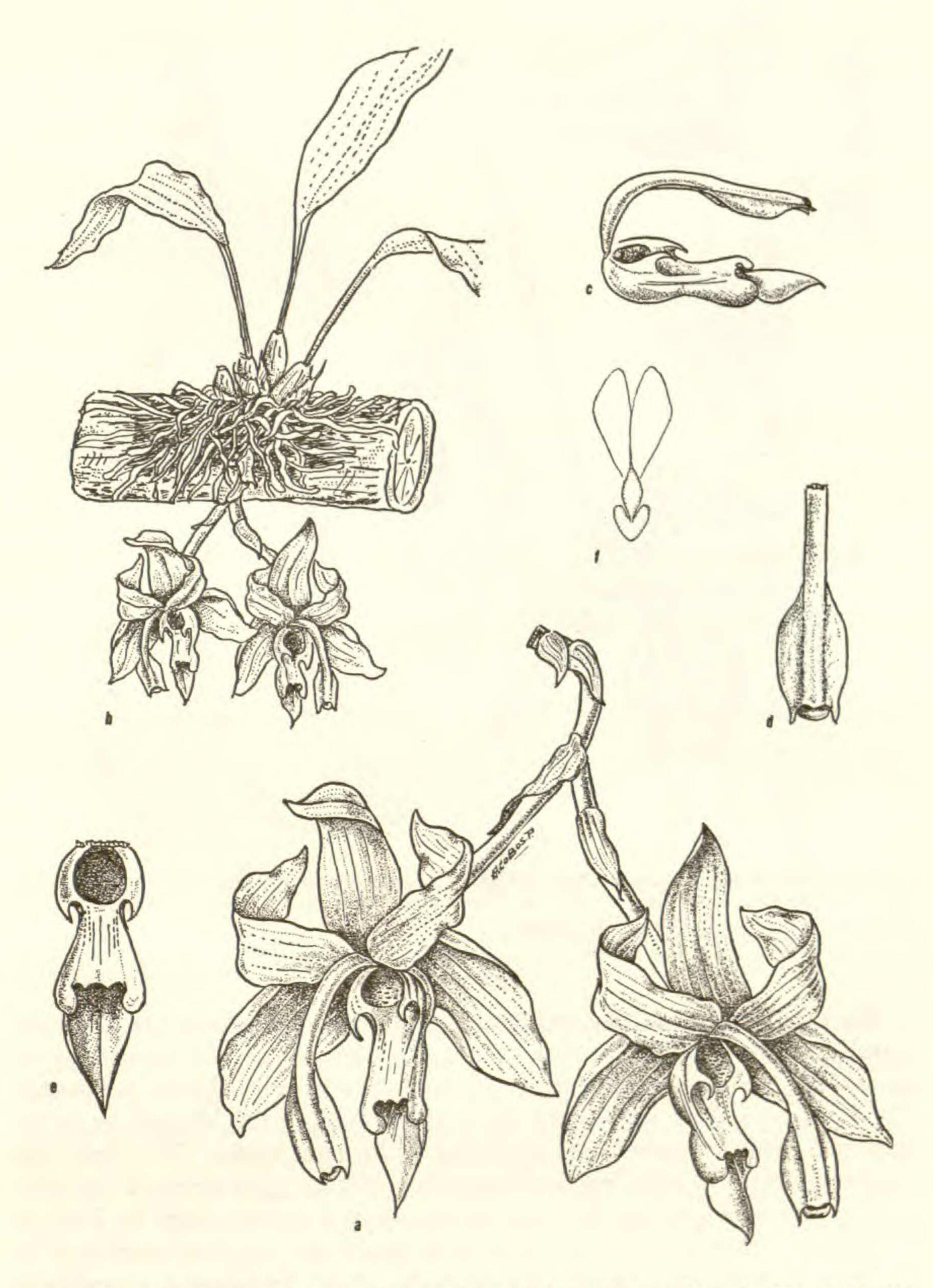


Fig. 11. Stanhopea grandiflora (Lodd.) Lindl. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{8})$. c. Side view of the lip $(\times \frac{1}{3})$. d. Dorsal view of the column $(\times \frac{1}{3})$. e. Dorsal view of the lip $(\times \frac{1}{3})$. f. Pollinarium $(\times 3)$.

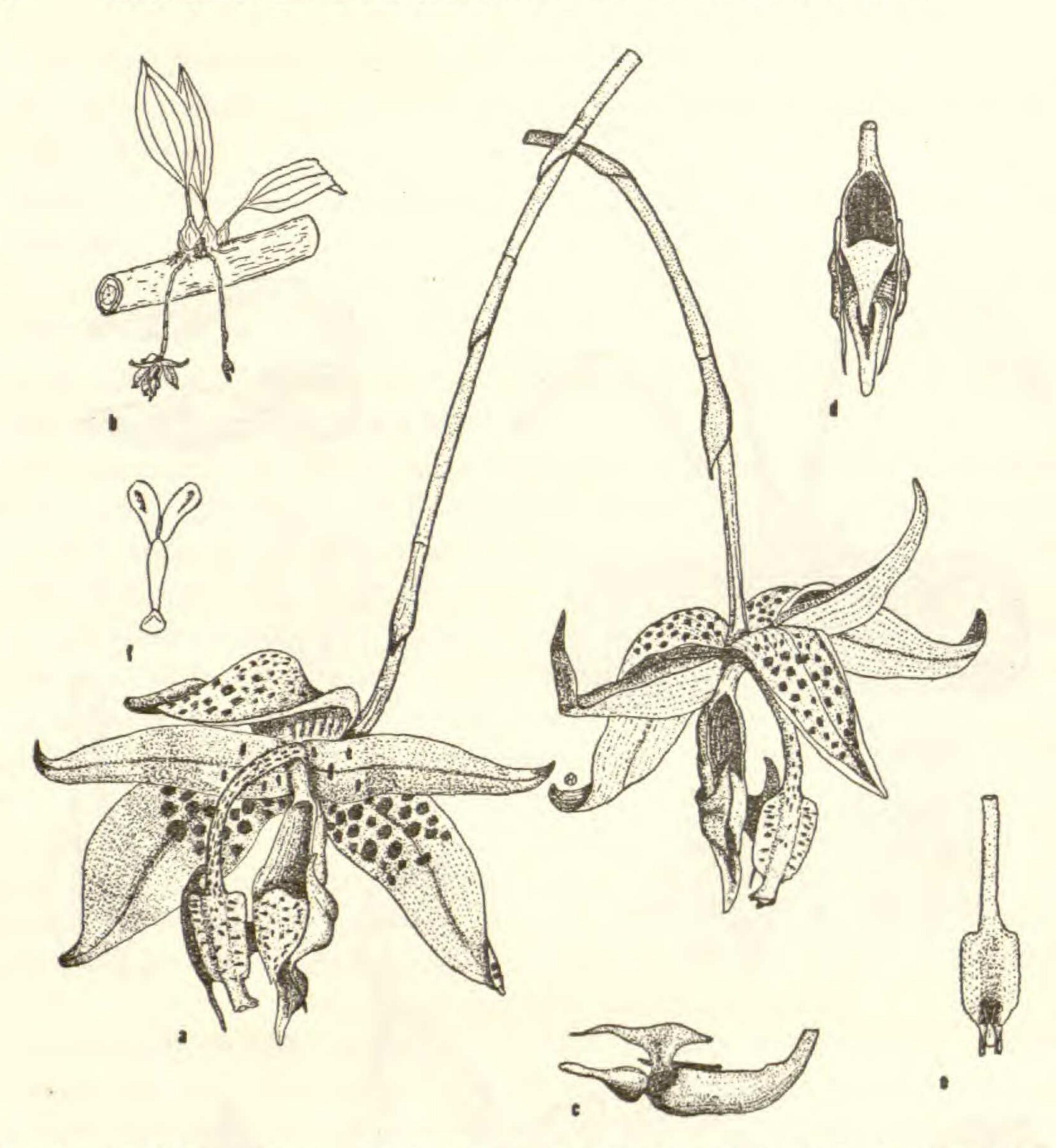


Fig. 12. Stanbopea rodigasiana Claes. ex Cogn. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{15})$. c. Side view of the lip $(\times \frac{1}{3})$. d. Dorsal view of the lip $(\times \frac{1}{3})$. e. Ventral view of the column $(\times \frac{1}{3})$. f. Pollinarium $(\times 2)$.

VARIATION

We have pointed out that populations of species in Stanhopea are often extremely variable. Unfortunately no extensive analysis of specific populations are yet available; however, we did superficially study two large populations in Ecuador. The first occurs along the western slopes of the Andes from Macuchi in northwestern Ecuador to Paccha in the south near the Peruvian border. This population is approximately 400 miles long and ranges from 400 to 1,200 meters in elevation.

Near Macuchi some of the plants correspond to a species named by Rolfe as S. anfracta. This species is very close to S. wardii and had been considered by Lindley as S. wardii var. venusta. Schweinfurth (1958) has placed it in synonymy with S. wardii. Other plants are similar to what has been known as S. bucephalus. The population was sampled at four locations through its central portion and was rather uniformly S. bucephalus with only occasional plants occurring which could be considered as referable to S. wardii. However, at Santa Isabela near the southern extension of the population the situation changed considerably. Plants were so

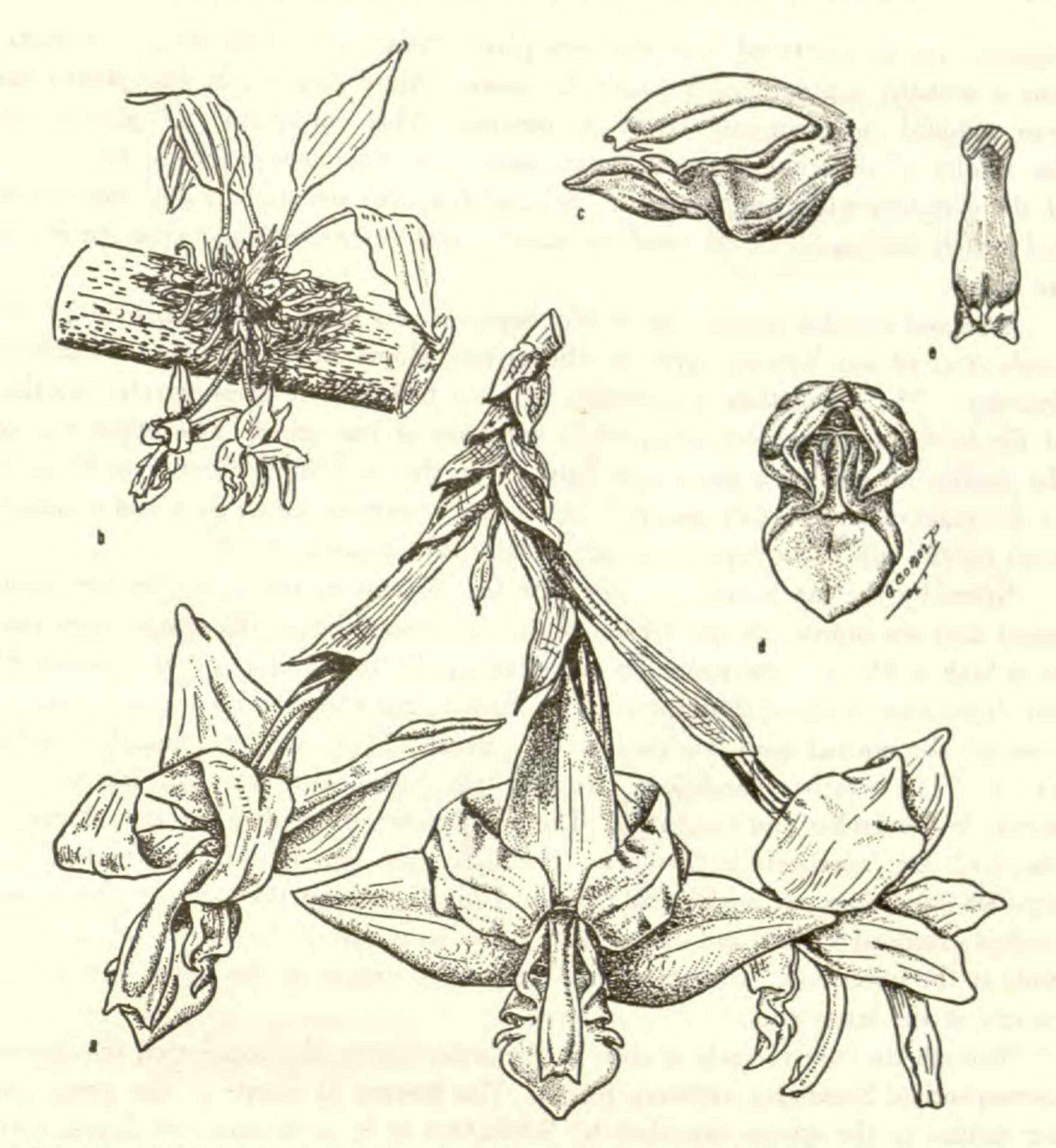


Fig. 13. Stanbopea lewisae Ames & Correll. a. Inflorescence $(\times \frac{1}{2})$. b. Plant habit $(\times \frac{1}{12})$. c. Side view of the lip $(\times \frac{2}{3})$. d. Dorsal view of the lip $(\times \frac{2}{3})$. e. Ventral view of the column $(\times \frac{2}{3})$.

variable that specimens were found which corresponded precisely to S. anfracta, S. bucephalus, S. graveolens, S. oculata, S. wardii, S. baseloviana, S. guttulata, S. ornatissima, S. platyceras, and S. peruviana. Many plants also occurred in the population which were intermediate between these concepts. Most of these species are actually color forms and many have been relegated to synonymy by other authors. Some of them have been maintained, largely because no botanist knew much about them, for they were described from individual specimens cultivated in Europe and their origins were unknown.

Plants from the population at Paccha, about 20 miles south of Santa Isabela were sent to Europe by a number of early collectors. Kruth named a plant which Humboldt and Bonpland had collected here as Epidendrum grandiflorum. Lindley later named another plant from the same locality as S. bucephalus and it was illustrated in the Botanical Register, plate 24, in 1845. Reichenbach changed the name to S. grandiflora, based on Kunth's name. Another illustration of a plant in the Botanical Magazine, Plate 5278 given as S. bucephalus is so inaccurate that four

"species" can be identified from the same plate. Plate 8517 in the Botanical Magazine is actually a plant much nearer S. wardii. More recently S. bucephalus has been reduced to synonymy under S. oculata. This confusion has pointed up the results of describing species from individual specimens without knowledge of the variation within populations. We feel that this situation is not uncommon and clearly demonstrates the need for careful and extensive population studies of the genus.

A second variable population of Stanhopea occurs in the valley formed by the headwaters of the Zamora river on the eastern slopes of the Andes in southern Ecuador. The population is extensive and we feel that it demonstrates another of the kinds of phenomena involved in variation of the genus. The upper end of the population reaches a point near Km. 45 on the road from Zamora to Loja, at an elevation of about 1800 meters. The valley is narrow, dense fogs and constant rains are the rule and temperatures are usually low, around 60° F.

Normally the sun breaks through only for an hour or two each day but occasional days are sunny. When the sun is out for several hours the temperature rises to as high as 75° F. The elevation decreases rapidly to the east and at Zamora, 62 km. from Loja, is about 1000 meters. At Zamora the valley levels out and becomes broader, the normal daily temperature is around 75° F. and occasionally reaches 85° F. The weather conditions change little during the year but less rainfall occurs in November and December. The population was studied as far as Yansasa, about 60 km. northeast of Zamora. Here the elevation is near 800 meters and weather conditions are similar to Zamora. In all, the portion of the population studied extended 85 km. and varied from 1 km. in width at the upper end to 3 km. wide at the lower end. Elevations run from 1800 meters at the upper end to 800 meters at the lower end.

Though they are variable in color at the lower end of the population the flowers correspond to Stanhopea anfracta Rolfe. The flowers of plants at the upper end are similar to the species described by Schlechter as S. peruviana and demonstrate little variation. Both of these species have been reduced to synonymy under S. wardii. A cline occurs from the upper end of the population to the lower, the plants in the center having flowers essentially intermediate between the extremes. Variation increases considerably toward the lower end of the population. Vegetatively the plants are indistinguishable throughout the population.

The flowers of the plants at the upper end of the population are morphologically nearly identical to the flowers at the opposite extremes but are colored quite differently and are much thicker in substance. The flowers are completely tangerine-yellow with no other color involved. The sepals and petals have heavy texture and are not reflexed. The flowers usually last for eight days.

At the lower end of the population the flowers are thin in substance and have their sepals and petals strongly reflexed. The basal part of the lip, sepals and petals are yellow-orange flecked and spotted with maroon-red, and the extremities of the sepals, petals and lip are white flecked with red. The flowers last only one or two days. Plants from around Zamora had flowers more or less intermediate between the two extremes in all characters.

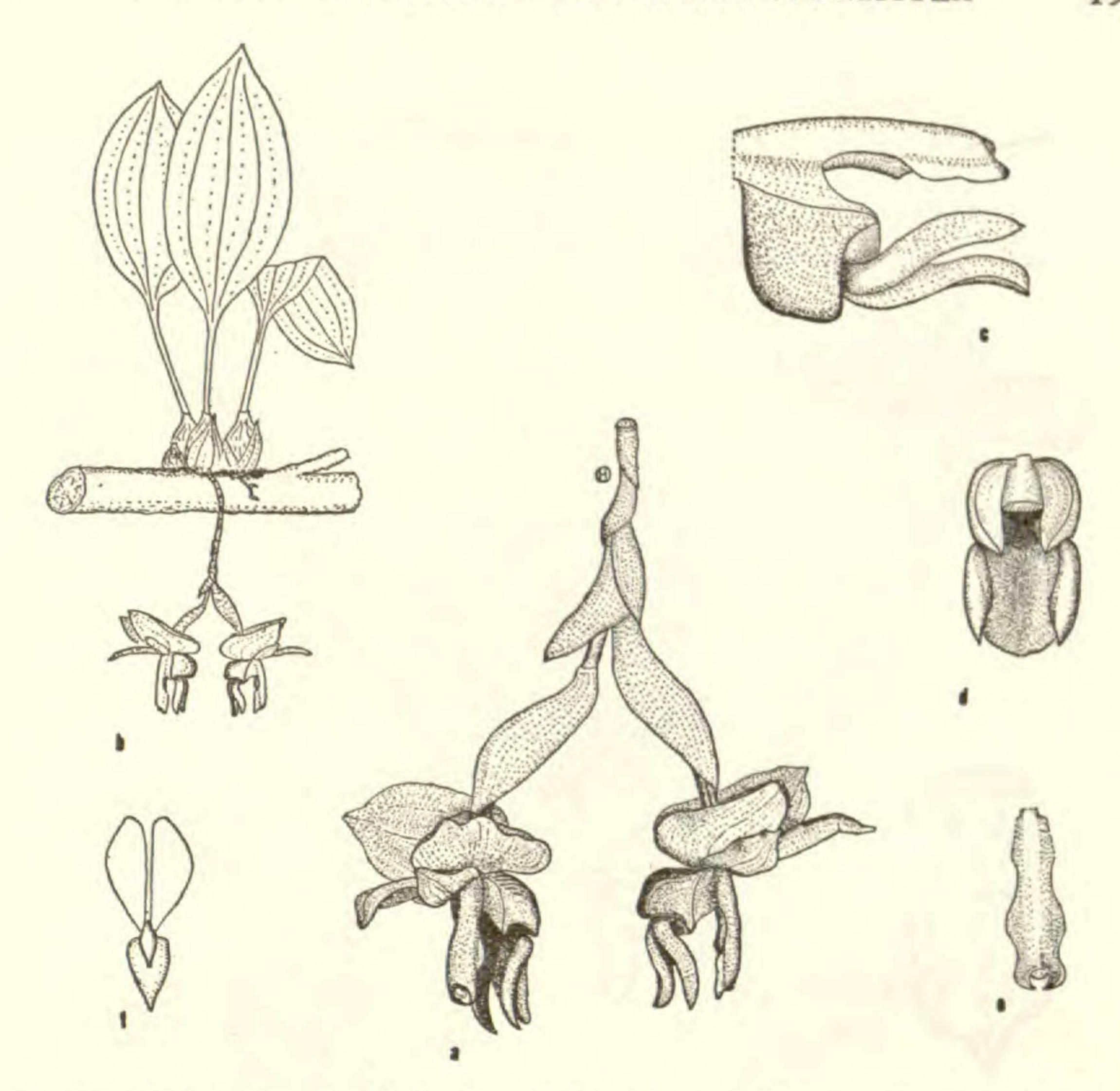


Fig. 14. Stanhopea connata Rchb. f. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{9})$. c. Side view of the lip $(\times \frac{2}{3})$. d. Dorsal view of the lip $(\times \frac{1}{2})$. e. Dorsal view of the column $(\times \frac{1}{2})$. F. Pollinarium $(\times 2)$.

From Zamora to Yansasa bees of the genus Eulaema were very common. One species in particular was quite common and appeared much like E. bomboides. They were of the same size but slightly different in color pattern and probably represent E. nivofasciata Friese. Though none was actually observed pollinating Stanhopeas it is probable that they are the pollinators. Eulaema cingulata was also common and while they visited other flowers placed in proximity they showed no interest in Stanhopeas and would not enter them.

The bees of this genus are partial to the warmer tropical climates. They nest in hollow limbs or in holes in embankments and their nests have not been encountered at elevations above 1,000 meters in Ecuador. From Zamora to Yansasa they are commonly observed visiting Costus, Inga and Bixa. They are fast flying bees and are capable of long flights. They are seldom seen in the upper end of the valley and then only on the rare, sunny, warm days. It would appear that they do not nest in the upper extensions of the valley. Plants in the lower regions commonly produce one or two seed capsules indicating considerable visitation by the insects. In the upper regions, on the other hand, seed capsules are rarely encountered.

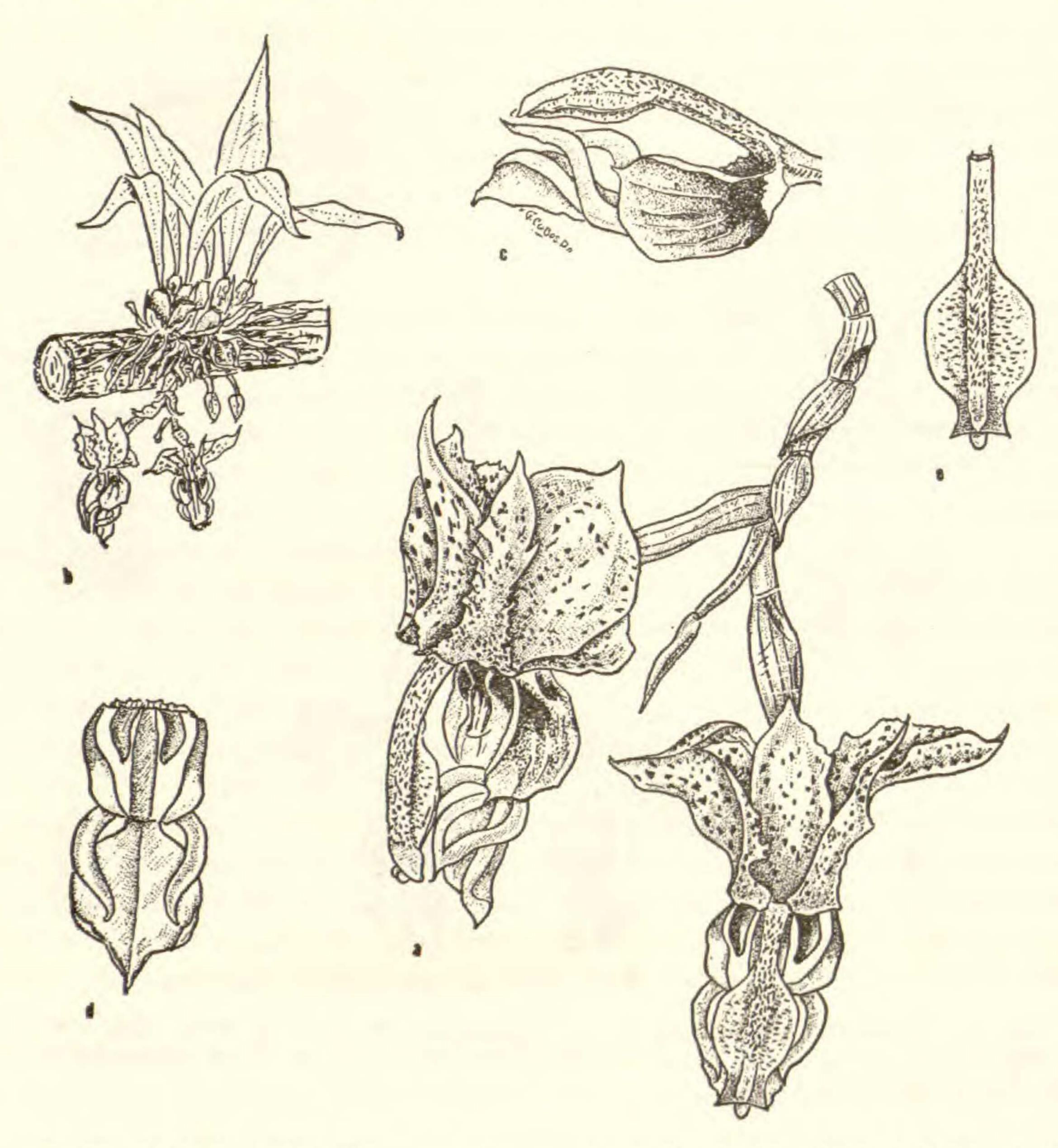


Fig. 15. Stanhopea insignis Frost. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{12})$. c. Side view of the lip $(\times \frac{1}{2})$. d. Dorsal view of the lip $(\times \frac{1}{2})$. e. Dorsal view of the column $(\times \frac{1}{2})$.

This population in some respects appears rather typical of the populations of Stanhopea in Ecuador. It is not as complex as the population of Stanhopea on the western slope of the Andes which is extremely variable in the direction of S. wardii in its northern portion, rather stable in its central portion corresponding to S. bucephalus and mixed with typical S. oculata in its southern extension. It is our opinion that this type of variation is typical of S. oculata and allied species throughout their range. It would appear that the relatively stable, heavy substanced, long lasting type of flower at the upper extension of the Zamora population may be due to strong natural selection for that type of flower. In the upper elevations where pollinators are not abundant it would be of more value to the population to have long lived flowers which would have a greater possibility of visitation and pollination. Such flowers may have to wait several days for weather conditions to be satisfactory for bees to reach them. Short lived flowers would stand little chance of being visited and therefore would rarely reproduce. Long life in these flowers depends on thickness of their parts. A selection pressure is therefore exerted for



Fig. 16. Stanhopea tigrina Batem. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{9})$. c. Side view of the lip $(\times \frac{1}{2})$. d. Dorsal view of the lip $(\times \frac{1}{3})$. e. Dorsal view of the column $(\times \frac{1}{2})$.

flowers with heavy substance. In the lower regions flowers are normally visited shortly after opening and long lived flowers would offer no selective advantage.

Some botanists would attempt to attribute such extreme population variability as is found in Stanhopea to hybridization and introgression. Perhaps this is a partial answer but in many cases the variable populations are so widespread as to preclude hybridization. One might also suggest that the variation is due to high mutation rate. Perhaps this is also a partial answer. It is apparent from what has been pointed out above in the pollination section of this paper that the bees which accomplish the pollination are attracted by the strong fragrance of the flowers. The color, and to a degree, the form of the flower appears to be of minor importance in its pollination. This is also true in the genera related to Stanhopea, such as Gongora and Coryanthes.

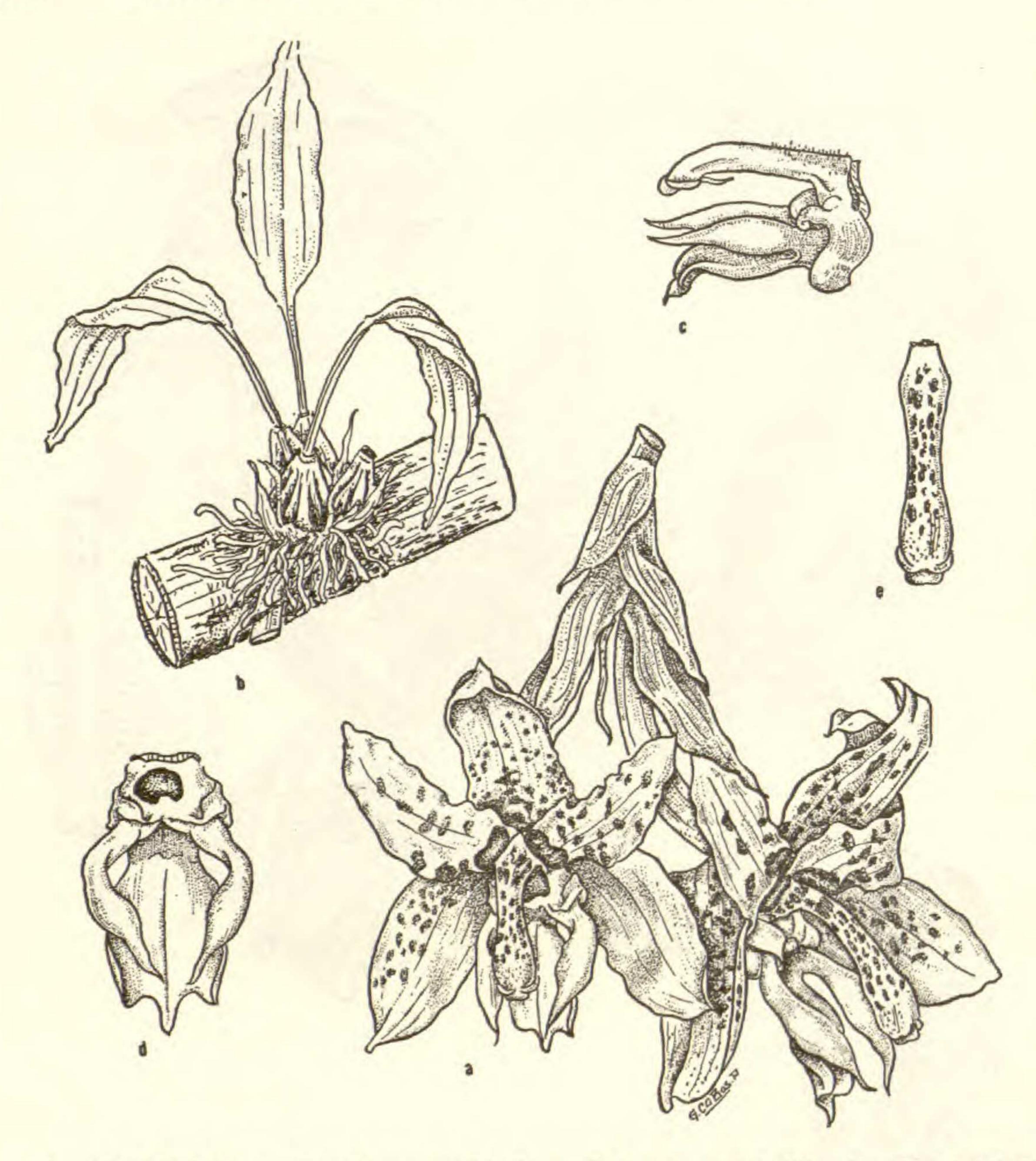


Fig. 17. Stanbopea martiana Batem. ex Lindl. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{9})$. c. Side view of the lip $(\times \frac{1}{2})$. d. Dorsal view of the lip $(\times \frac{1}{2})$. e. Dorsal view of the column $(\times \frac{1}{2})$.

If the attraction of the fragrance is so strong, color may be a minor factor in selection and the bees will still visit the flowers even if color varies widely. Also form could vary widely in parts which are not essential to the actual mechanisms of pollination, allowing variants due to mutation to remain within the population as fully effective breeding members contributing to the gene pool as a whole. In S. oculata and its allied species the characters which vary the most are the form of the hypochile of the lip and the color of the lip, sepals and petals. These are characters which are not actually involved in the mechanisms of pollination, and may not be important in the actual attraction of the pollinator. Selection would not be strong for these characters and a plant which deviated from the norm to a considerable extent could still remain a breeding member of the population as long as it retained the character of fragrance to attract the pollinator and the correct propor-



Fig. 18. Stanhopea saccata Batem. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{5})$. c. Side view of the lip $(\times \frac{1}{2})$. d. Dorsal view of the lip $(\times \frac{1}{2})$. e. Dorsal view of the column $(\times \frac{1}{2})$.

tions of the tunnel of the lip, formed by the mesochile and column, to properly place the pollinia for transport by the bee to another flower.

The fact that cross-pollination is nearly obligate in these genera may be of importance also. Certainly more variation would be produced, maintained and dispersed in a population of obligate cross-pollinating plants than in a population where self-pollination is possible and often of regular occurrence. Certain of the genera in the Catasetinae, i.e., Catasetum and Cycnoches, are characterized by unisexual flowers which make cross-pollination obligate. These are genera in which the species are notoriously variable. In many of the other genera self-

incompatibility is the rule. Not a great deal of information is available on this phenomenon but the authors have made numerous attempts to self-pollinate Stanhopea bucephalus and Gongora maculata with no success. The resulting seed pods dropped after a few weeks.

In most of these variable genera the flowers are highly fragrant but extremely short lived, seldom lasting more than two or three days. The fragrance ceases very soon after the flower has been pollinated. It would appear that there is a correlation involved between the characteristics of powerful fragrance and short lived flowers. These species seem to depend on their fragrance to attract pollinators and are usually very successful, for a considerable portion of the flowers are pollinated. They have developed in the direction of stronger and more attractive fragrances but offer no food or nectar to the pollinator. Selection may even favor short-lived flowers which do not attract bees long after their pollination (i.e., do not compete with the younger flowers). Here there seems to be a strong selection pressure for fragrance for pollinator attraction with lower selection pressure for constancy of morphological characters.

Many of the orchids which have long lasting flowers are either not fragrant or are weakly fragrant. These orchids seem to depend on numbers of flowers few of which are ever pollinated. These species for the most part are not as variable as the fragrant types and have developed other mechanisms for attraction of the pollinator. Where populations of these species are extremely variable, it is usually in a relatively limited geographical area and their variability can often be attributed to introgressive hybridization.

SUCCESSFULNESS OF THE GENUS

The objective measurement of success in a group of plants is difficult, as too many factors are involved. Obviously, so called weedy species would rate high on the scale. However, to attempt to rate other plants as being highly successful or only fairly successful is scarcely possible. We do feel, however, that we can subjectively rate the success of a group of orchids in relation to other orchids. In such a subjective sense we can use extent of distribution and density of populations as measuring tools. Many orchids are highly successful in respect to density of population but are limited in distribution. Others have wide distributions but are seldom encountered in quantity in a given area. The majority of the species of Stanhopea are in one or the other of these categories. The "insignis and oculata complexes" on this basis are highly successful in that they are commonly found in quantity (in undisturbed habitats) and have broad ranges. They are to be encountered in any region of the American tropics which provides the basic needs of the plants. They are found from sea level to 7000 feet and grow as well in either location provided there exists sufficient humidity and shade and the temperatures are not too low.

This very success, we feel, contributes to their variability, but on the other hand, may have suppressed speciation. The extremes of the range may be expected to produce forms which are unlike due simply to spatial relations. However isolation is difficult to achieve in a dense and essentially continuous population. In

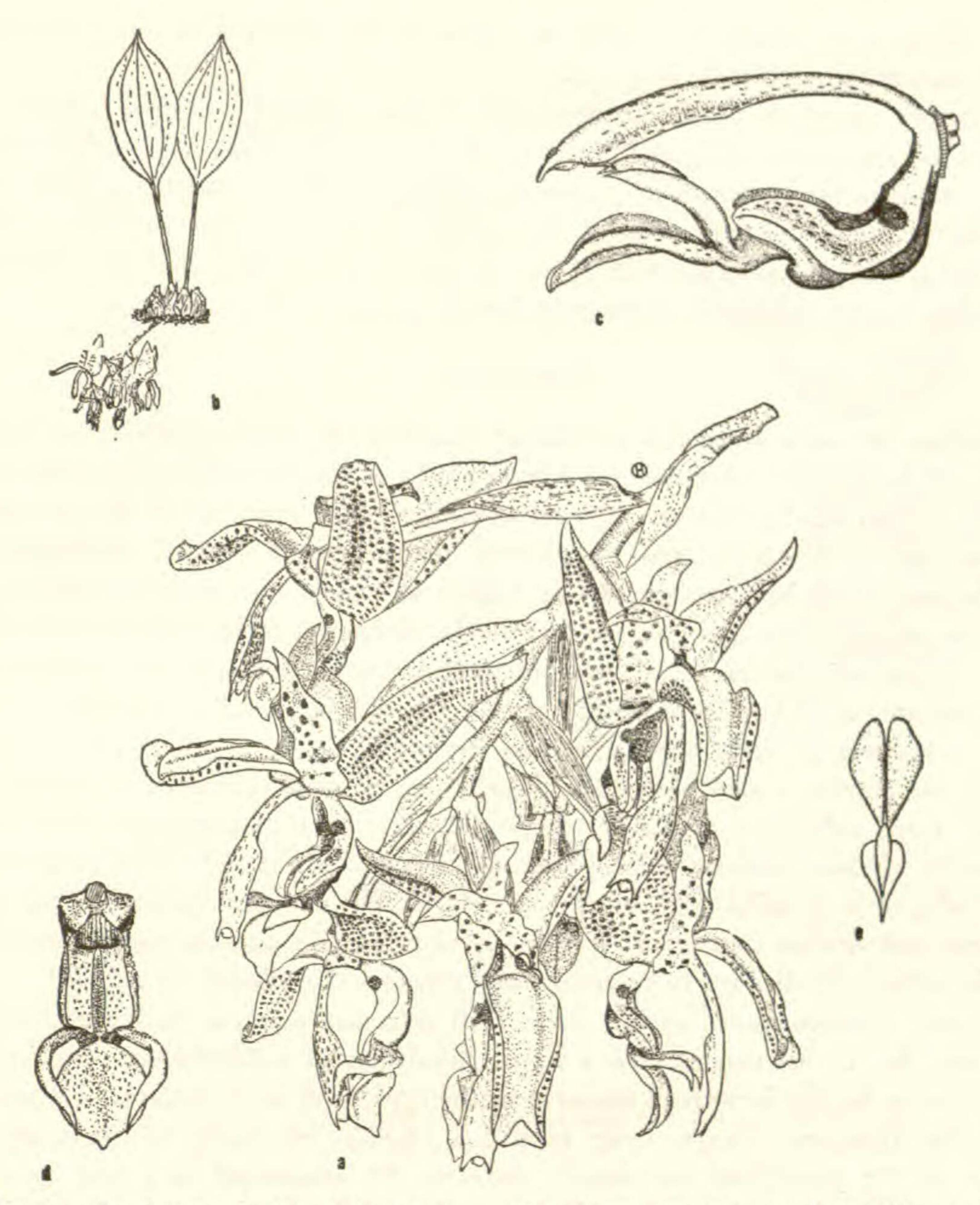


Fig. 19. Stanhopea oculata (Lodd.) Lindl. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{15})$. c. Side view of the lip $(\times \frac{2}{3})$. d. Dorsal view of the lip $(\times \frac{1}{2})$. e. Pollinarium $(\times 2)$.

this respect "success" and taxonomic diversity may be mutually incompatible. This situation is by no means limited to Stanhopea. The hooded types of Catasetum reflect the same phenomena, though not quite to the same degree. In Mexico and middle Central America a form known as C. integerrimum Hook. is common and variable. Further south in Costa Rica and Panama intergrading populations known as C. viridiflavum Hook., C. oerstedii Rchb. f. and C. maculatum Kunth occur. From Panama extending through Venezuela, the Guianas and northern Brazil the form known as C. macrocarpum L. C. Rich. is found. In Ecuador C. macroglossum Rchb. f. is common. All of these populations are highly variable and in Ecuador alone, specimens corresponding to all of these "species" may be found. There is little reason to recognize these species when they are now known to be only representatives of one widely distributed and highly successful popula-

tion. Here, as in Stanhopea, speciation seems to be inhibited by the otherwise highly successful population structure.

Gongora maculata Lindl. demonstrates a close parallel. Several species of Cycnoches, Mormodes, Coryanthes, Epidendrum and Oncidium have similar patterns. Many other instances of highly successful, widely distributed orchids can be given in which the most striking chracteristic is their extreme variability. We feel that speciation has taken a back seat to success in the sense that development of distinct taxa is inhibited where gene flow is unrestricted and selection is weak.

TAXONOMY

Perhaps the most successful method of handling the great variability in Stan-hopea will be to follow that used by Allen (1952) in his particularly workable and useful revision of Cycnoches. Cycnoches reflects the same kinds of problems encountered in Stanhopea and by placing several closely related, intergrading populations, which had been previously known as species, as subspecies under one inclusive species, Allen was able to indicate the close relationships of these populations. In our opinion this is the most practicable manner to handle the problems of such extreme variability in obviously closely related populations. Certainly, it is folly to attempt to apply species or even subspecies names to all of the variants found; the number would be endless. There may be some difference of opinion as to how many subspecies should be recognized, some authors suggesting more and others less. When numerous well authenticated population studies are accomplished in Stanhopea it is quite possible that certain of the dubious species within the "insignis and oculata complexes" will be found to exist as discrete populations and will be sufficiently distinct to be recognized, others will probably be reduced.

Recent correspondence with a rather well informed amateur orchid enthusiast indicates that he has encountered a stable population of a Stanhopea. Specimens which he has kindly forwarded appear near the type form of S. oculata. He maintains that these are different from typical S. oculata, are stable within what he knows of the population and should therefore be recognized as a new species. Perhaps with exhaustive study this population might be found to be distinct throughout its extent and should therefore be considered as a valid species. Our experience with populations of Stanhopea in Mexico, Guatemala, Costa Rica, Panama, Ecuador and Peru would indicate that this population, throughout its entire range, would very doubtfully represent even a valid subspecies of S. oculata. This case, however, is quite representative of the problems which Stanhopea offers and indicates the necessity of thorough knowledge of large percentages of over-all populations before rash action is taken in applying more epithets in the group.

The nomenclature of Stanhopea is badly confused and the efforts made by modern workers to clarify the situation have not always been particularly successful. Many of the early descriptions of the species were not accompanied by illustrations and were at best brief. Several of them are nearly impossible to identify with any particular population. One of the unfortunate pitfalls the modern taxonomist encounters is the tendency to make one of these early descriptions fit his concept of a given species or vice versa. Normally the taxonomist has the type specimen

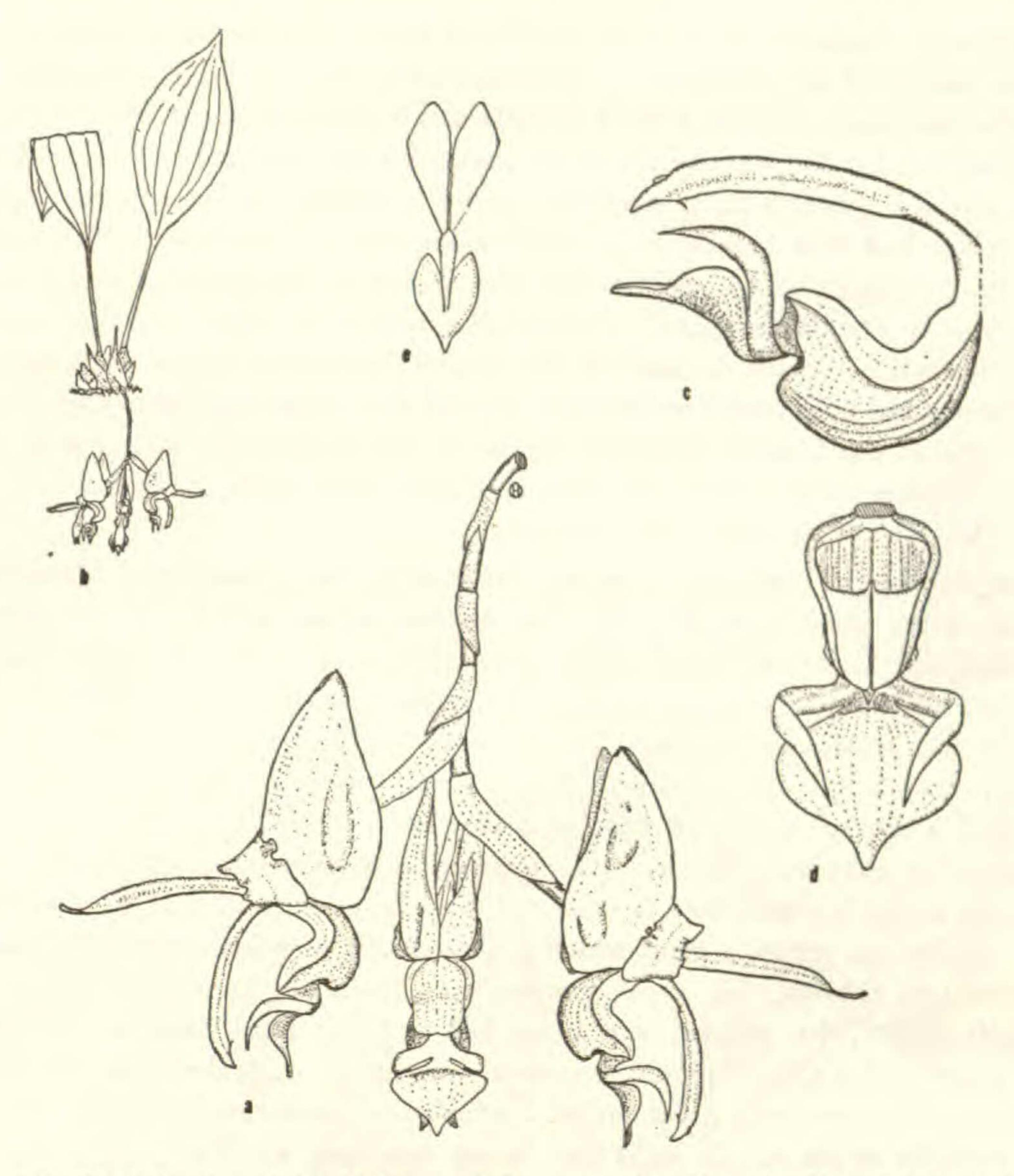


Fig. 20. Stanhopea bucephalus Lindl. a. Inflorescence (X 1/3). b. Plant habit (X 1/15). c. Side view of the lip (X 2/3). d. Dorsal view of the lip (X 2/3). e. Pollinarium (X 2).

to fall back on and this specimen, when carefully examined, supplies the necessary information to avoid error. Unfortunately, herbarium specimens of Stanhopeas do not always fulfill this function even when they exist. The critical characters which separate species in this genus are for the most part destroyed in the preparation of specimens.

Many times the type specimen does not reflect the actual structure of the population. An example is to be found in S. wardii in which the type description mentions two teeth produced at the base of the hypochile by the extension of the falcate lateral margins. This character is common in the specimens from southern Central America and perhaps to a lesser extent in Guatemala, but actually does not occur in many of the specimens from Mexico, Nicaragua and northern South America, which are certainly referable to this species on every other criterion. To further complicate the situation, this character does occur occasionally in other species such as S. oculata and S. bucephalus. Doubtless, the type specimen of S. wardii did have this character, but the presence or absence of this one character,

since it is not diagnostic of the total population and is found in other species, should not be considered as a criterion for determination. Several other characters help to make this species distinct from S. oculata and S. bucephalus. Allen (1949) has suggested that Lindley was in error in his description of S. oculata and S. bucephalus. Allen placed S. bucephalus in synonymy under S. oculata and re-erected S. graveolens which had been considered as synonymous with S. bucephalus. We cannot, however, agree with Allen and feel that the epithet, S. bucephalus, should be used in place of S. graveolens. Since Lindley's descriptions are extremely brief and the later illustrations which he used of his species correspond rather well with his descriptions and with actual populations, we feel that Allen has allowed his splendid knowledge of the Central American species to overshadow the variation of these species in other areas. Since the type specimens yield little information it will always be difficult to resolve this situation.

Another of the confusing situations involved in the taxonomy of Stanhopea is the use of the name "grandiflora". This name was first used by Kunth when he described and illustrated Epidendrum grandiflorum in 1808. In 1828 Loddiges described another plant and named it Ceratochilus grandiflorus but the name Ceratochilus had previously been used by Blume for an East Indian orchid. In 1832 Lindley transferred the Loddiges plant to the genus Stanhopea of Frost and described a similar species as Stanhopea eburnea. Later these two plants were found to be conspecific but have been known as S. eburnea to the present time. Lindley's name, S. grandiflora, is valid and must be used in place of S. eburnea. In 1832 Lindley also named a plant which was essentially identical to Kunth's plant as Stanhopea bucephalus. The type specimen was a Ruiz and Pavón collection which, as Rolfe (1912) has pointed out, is marked as being from Mexico. It appears that many of Ruíz and Pavón's specimens were mixed with the Sessé and Mociño Mexican collections and a great deal of confusion has been caused by this situation. The probable origin of the Ruíz and Pavón specimen was Peru, as it was then called, or what is now Ecuador. It must have come from very near the same locality as Kunth's specimen since the trails of access to the Andes were very few during this period.

Reichenbach apparently attempted to clear up the situation by transferring Kunth's plant to Stanhopea and reducing Lindley's S. bucephalus to synonymy under S. grandiflora Rchb. f. Unfortunately, Reichenbach's S. grandiflora is invalid because Lindley's combination precedes it.

Another attempt to clarify the situation in the "insignis complex" has been the reduction of the name S. tigrina Batem. to synonymy under S. devoniensis Lindl. by Williams (1951). However, the concept, early descriptions, and illustrations of S. devoniensis and S. tigrina appear to us to be clearly distinct. The plate of S. devoniensis in Lindley's Sertum Orchidacearum has little resemblance to Bateman's plate of S. tigrina in his Orchids of Mexico and Guatemala and very likely represents either a rarely encountered species or a hybrid form between S. saccata and S. tigrina. The figure very definitely resembles S. saccata in the features of the lip more closely than it does S. tigrina. Williams may have been misled by the color, and was certainly handicapped by the lack of good type material. Occasionally

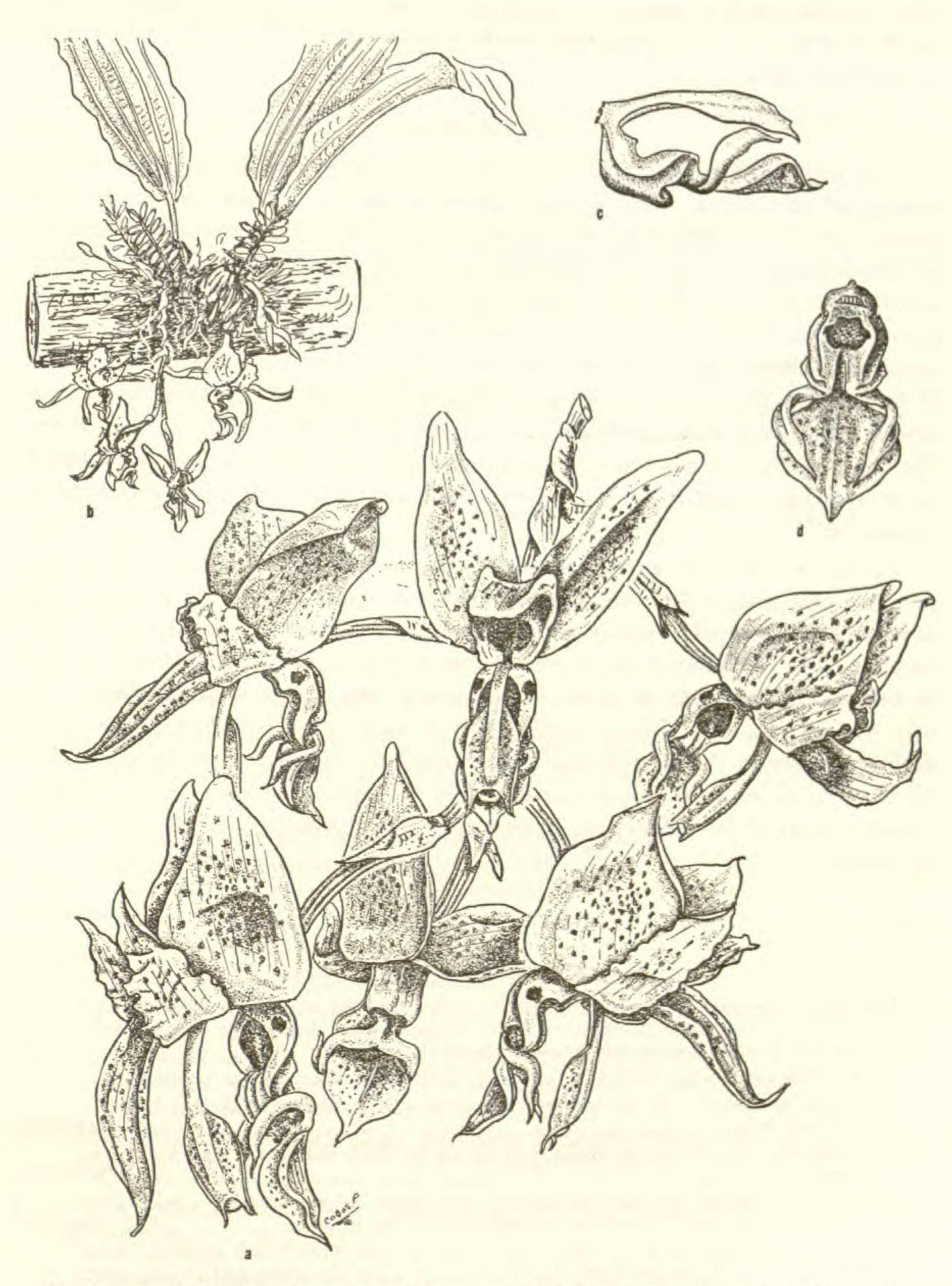


Fig. 21. Stanhopea wardii Lindl. a. Inflorescence $(\times \frac{1}{3})$. b. Plant habit $(\times \frac{1}{9})$. c. Side veiw of the lip $(\times \frac{1}{2})$. d. Dorsal view of the lip $(\times \frac{1}{2})$.

plants with flowers somewhat like the description and plate (discounting the three-lobed epichile which is lacking in the plate but mentioned in the description) turn up, but are quite rare in comparison to the number of typical S. tigrina found in the Mexican populations.

CONCLUSIONS

This paper has been written as a discussion of Stanhopea preliminary to a formal revision of the genus. Our original intention was to include the revision of Stanhopea with this paper but after examination of herbarium material proved to be disappointing we have concluded that only with further field study of living populations can we accomplish a truly meaningful treatment of the genus. We have also found that to merely revise Stanhopea and leave the other members of the subtribe Stanhopeinae in the condition which they are at the present time would be to do a less than meaningful work. The members of the Stanhopeinae are so closely intermeshed with Stanhopea that revision of the whole group is mandatory. We hope that in the not too distant future such a revision can be accomplished using classical methods of taxonomy combined with modern biosystematic procedures.

In reality then, this paper can only point up the problems involved in the group; it brings up to date the information that is available from the literature and adds our own limited observations of the natural phenomena occurring in the group. These field observations have pointed out the path, so to speak, that should be followed in the study of such a complex and difficult genus as Stanhopea. It may well be true that similar procedures will need to be used in treating other problem groups in the Orchidaceae. Our experience indicates that future studies of the difficult subtribe Catasetinae can only be approached through field study. Probably parts of the Sobraliinae, Epidendrinae and Pleurothallidinae will prove to be similar.

APPENDIX

KEY TO STANHOPEA

- a. Lip simple, composed of a saccate hypochile with thickened midlobe forming a nonarticulated epichile, mesochile absent; column thick, wingless or narrowly winged with the apex of the wings caudate and extended beyond the anther.
 - b. Hypochile entire, not projected into side lobes on its upper edges; column wingless.
- aa. Lip complex, composed of 2 or 3 distinct parts, epichile articulated to the hypochile or to a fleshy mesochile that is provided with lateral horns; column slender and arcuate with narrow or broad wings which are not caudate at their apex and extended beyond the anther.

 - bb. Petals thin in substance and reflexed; mesochile of the lip with 2 horns on its margins or none.

- c. Hypochile of the lip provided with 2 distinct horns or protuberences, mesochile relatively simple and without lateral horns.
- cc. Hypochile of the lip with the lateral margins thickened, not provided with horns or protuberences, mesochile with 2 distinct horns.
 - d. Horns of the mesochile divided at their midpoint to produce a falcate, retrorse appendage as well as a slender forward projecting spine; flowers always solitary.

 7. S. RODIGASIANA
 - dd. Horns of the mesochile acicular or flat, not divided; flowers normally 2 or more.

 - ee. Horns of the mesochile long, free for more than 2 centimeters.
 - f. Mesochile obsolete, very short, hypochile conspicuously adnate to the column.

 9. S. CONNATA
 - ff. Mesochile conspicuous, well developed, hypochile not adnate to the column.

LIST OF SPECIES*

- 1. STANHOPEA ECORNUTA Lem. in Fl. des Serres 2: t. 181. 1846. (Fig. 6).

 Stanhopea calceolus Hort. ex Rchb. f. Xen. Orch. 1:117. 1858.

 Stanhopeastrum ecornutum Rchb. f. Xen. Orch. 1:124. 1858.
- 2. STANHOPEA PULLA Rchb. f. in Gard. Chron. 7:810. 1877. (Fig. 7).
- 3. STANHOPEA CIRRHATA Lindl. in Journ. Hort. Soc. 5:37. 1850. (Fig. 8).
- 4. STANHOPEA TRICORNIS Lindl. in Journ. Hort. Soc. 4:236. 1849. (Fig. 9).

†Stanhopea stenochila Lehm. & Kranzl. in Gard. Chron. 369. 1900.

†Stanhopea langlasseana Cogn. in Gard. Chron. 426. 1901.

Stanhopea convulata Rolfe, in Kew Bull. 366. 1909.

STANHOPEA REICHENBACHIANA Roezl. ex Rchb. f. in Gard. Chron. 2:40. 1879.
 (Fig. 10).

Stanhopea amesiana Hort. ex Gard. Chron. 352. 1893.

†Stanhopea lowii Rolfe, in Kew Bull. 63. 1893.

Stanhopea suavis Hort. in Ospina, Orquidas Colomb.

6. STANHOPEA GRANDIFLORA (Lodd.) Lindl. Gen. & Sp. Orch. 158. 1832. (Fig. 11).

Ceratochilus grandiflorus Lodd. in Bot. Cab. t. 1414. 1828.

Stanhopea eburnea Lindl. in Bot. Reg. 18: t. 1529. 1832.

Stanhopea calceolata Hort. ex Rchb. f. 1:117. 1858.

Stanhopea calceolata Drap. Leng. Hort. Univ. 2:127 & 264.

†Stanhopea candida Barb. Rodr. Gen. Spec. Orch. Nov. 1:101. 1877.

Stanhopea randii Rolfe, in Kew Bull. 363. 1894.

- 7. STANHOPEA RODIGASIANA Claes. ex Cogn. in Gard. Chron. 14. Fig. 9. 1898. (Fig. 12).
- 8. STANHOPEA LEWISAE Ames & Correll, in Bot. Mus. Leafl. Harv. Univ. 104:86. Pl. 10. 1942. (Fig. 13).
- 9. STANHOPEA CONNATA Kl. in Otto & Deitz. Alleg. Gartenz. 22:226. 1854. (Fig. 14).

Stanbopea graveolens Kl. ex Rchb. f. Xen. Orch. 1:118. 1879

^{*} The names placed in synonymy in this list have been previously reduced by other authors with the exception of those marked thusly: †.

10. "insignis complex"

STANHOPEA INSIGNIS Frost ex Hook. in Bot. Mag. t. 1948 & 1949. 1829. (Fig. 15).

Stanhopea flava Lodd. ex Beer. Prakt. Stud. Fam. Orch. 312. 1854.

Stanhopea atropurpurea Lodd. ex Planch. Hort. Donat. Orch. 215. 1858.

Stanhopea odoratissima Hort. ex Planch. Hort. Donat. Orch. 216. 1858.

Stanhopea macrochila Lem. in Illustr. Hort. 4: Misc. 71. 1859.

STANHOPEA TIGRINA Batem. ex Lindl. Sert. Orch. 1. 1838. (Fig. 16).

Stanhopea cavendishii Lindl. ex Batem. Lond. Hort. Brit. Suppl. 3:643.

Stanhopea maculosa Knowles & Westc. Flor. Cab. 3: t. 121. 1839.

Stanhopea nigroviolacea Morren. ex Beer. Prakt. Orch. 313. 1854.

Stanhopea expansa P. N. Don. Hort. Cantab. 13:721. 1845.

Stanbopea lyncea P. N. Don. Hort. Cantab. 13:608. 1845.

STANHOPEA DEVONIENSIS Lindl. Sert. Orch. t. I. 1838.

STANHOPEA MARTIANA Batem. ex Lindl. in Bot. Reg. 26: Misc. 50. 1840. (Fig. 17).

Stanbopea velata Morren, in Ann. Soc. Gand. 3. t. 153. 1847.

STANHOPEA SACCATA Batem. Orch. Mex. & Guatem. t. 15. 1839.

Stanhopea implicata Westc. ex Lindl. Fol. Orch. Stan. 6. 1852.

Stanhopea radiosa Lem. Illustr. Hort. 4: Misc. 72. 1859.

Stanhopea marshii Rchb. f. Xen. Orch. 1:120. 1879.

Stanhopea elegantula Rolfe, in Kew Bull. 161. 1910.

STANHOPEA INTERMEDIA Kl. in Act. Hort. Petrop. 17:142. 1898.

11. "oculata complex"

STANHOPEA OCULATA (Lodd.) Lindl. Gen. & Sp. Orch. Pl. 158. 1832. (Fig. 19).

Ceratochilus oculatus Lodd. in Bot. Cab. t. 1764. 1832.

Stanhopea lindleyi Zuccar, Abh. Akad. Muenich 2:320. 1831-33.

Stanhopea guttulata Lindl. in Bot. Reg. 29: Misc. 75. 1843.

Stanhopea graveolens Morren in Ann. de Gard. 2:55. t. 54. 1846.

Stanhopea guttata Kich. Berl. Allg. Gartenz. 364. 1858.

Stanhopea ornatissima Lem. in Illustr. Hort. 9:325. 1862.

Stanhopea cymbiformis Rchb. f. Xen. Orch. 2:84. t. 124. 1865.

Stanhopea minor Schltr. Notizbl. Bot. Gart. Mus. Berlin 6:483. 1917.

STANHOPEA BUCEPHALUS Lindl. Gen. & Sp. Orch. Pl. 157. 1832. (Fig. 20).

Stanhopea graveolens Lindl. in Bot. Reg. 3: Misc. 59. 1840.

Stanhopea aurantia Lodd. ex P. N. Don in Hort. Cantab. 608. 1845.

Stanhopea jenishiana Kramer ex Rchb. f. Bot. Zeit. 10:934. 1852.

Stanhopea warscewicziana Kl. in Allg. Gartenz. 20:214. 1852.

Stanhopea guttata Beer. Prakt. Orch. 312. 1854.

Stanhopea inodora Rchb. f. Xen. Orch. 2:157. 1858.

Stanbopea aurata Hort. ex Planch. Hort. Donat. Orch. 104. 1858.

Stanhopea aurata Beer, Prakt. Stud. Fam. Orch. 310. 1858.

Stanbopea costaricensis Rchb. f. in Hamb. Gartenz. 16:424. 1860.

Stanbopea grandiflora Rchb. f. Walp. Ann. 6:587. 1863.

Stanhopea oculata var. constricta Kl. in Acta Hort. Petrop. 17:15. 1898.

Stanhopea lietzei Schltr. in Fedde Rep. Sp. Nov. 16:248. 1919.

Stanhopea remota Hort.

STANHOPEA WARDII Lodd. ex Lindl. Sert. Orch. t. 20. 1838. (Fig. 21).

Stanhopea aurea Lodd. ex Lindl. in Bot. Reg. 4: Misc. 11. 1841.

Stanhopea venusta Lindl. in Bot. Reg. 4: Misc. 11. 1841.

Stanbopea inodora Lodd. ex Lindl. in Bot. Reg. t. 65. 1845.

Stanhopea amoena Kl. in Allg. Gartenz. 20:273. 1852.

Stanbopea anfracta Rolfe, in Orch. Rev. 12:357. 1904.

Stanhopea purpusii Schltr. in Orchis 10:186. 1916.

Stanhopea peruviana Rolfe, in Bot. Mag. t. 8417. 1912.

SPECIES TOO POORLY KNOWN TO PLACE AT PRESENT

STANHOPEA ANNULATA Mansf. in Orchis 16:19. 1938.

STANHOPEA BICOLOR C. Koch. Berl. Allg. Gartenz. 209. 1857.

STANHOPEA DELTOIDEA Lem. in Illustr. Hort. 9:340. 1862.

STANHOPEA FLORIDA Rchb. f. in Gard. Chron. 615. 1879.

STANHOPEA FREGEANA Rchb. f. in Otto & Dietr. Allg. Gartenz. 23:313. 1855.

STANHOPEA FUERSTENBERGIAE Kranzl. in Gard. Chron. 161. 1899.

STANHOPEA GIBBOSA Rchb. f. in Gard. Chron. 1254. 1869.

STANHOPEA HASELOVIANA Rchb. f. in Otto & Dietr. Alg. Gartenz. 23:322. 1855.

STANHOPEA HOPPII Schltr. in Fedde Rep. Sp. Nov. 27:82. 1924.

STANHOPEA IMPRESSA Rolfe, in Kew Bull. 196. 1898.

STANHOPEA MADOUXIANA Cogn. in Gard. Chron. 134. 1898.

STANHOPEA MOLIANA Rolfe, in Lindenia 7:89. t. 331. 1891.

STANHOPEA NIGRIPES Rolfe, in Kew Bull. 364. 1894.

STANHOPEA PLATYCERAS Rchb. f. in Gard. Chron. 27. 1868.

STANHOPEA RUCKERI Lindl. in Bot. Reg. subt. 44. 1843.

STANHOPEA SCHILLERIANA Rchb. f. Xen. Orch. 2:158. 1858.

STANHOPEA SHUTTLEWORTHII Rchb. f. in Gard. Chron. 795. 1876.

STANHOPEA UNCINATA Drap. Hort. Univ. 4:65. 1843.

STANHOPEA XYTRIOPHORA Rchb. f. in Gard. Chron. 842. 1868.

DUBIOUS SPECIES

Stanhopea quadricornis Lindl. in Bot. Reg. 24: t. 5. 1838.

OBSCURE SPECIES

Stanhopea russelliana Lodd. ex P. N. Don in Donn. Hort. Cantab. 13:721. 1845. Stanhopea violacea Hort. ex Beer. Beitr. Morph. u Biol. Orch. t. 4 & 8. 1863. Stanhopea wallisii Rchb. f. in Linnaea 41:109. 1877.

Stanhopea hernandezii (Kunth) Schlechter, Beih. Bot. Centr. 2:490. 1918.

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EXPLANATION OF PLATE

PLATE III

Pollination in Stanbopea

A. Eulaema bomboides attempting to land on the hypochile of Stanhopea bucephalus. B. Bee falling through the flower and engaging the spine of the rostellum under his metathorax. C. Pollinia being inserted into the stigmatic cleft while the bee is falling through the flower. D. Eulaema moriana approaching the flower of S. tricornis.